



Joint CASP16-CAPRI57 assembly prediction round of 2024

Marc Lensink, Sameer Velankar, Alexandre Bonvin

University of Lille, CNRS, France
European Bioinformatics Institute (EMBL-EBI), Hinxton, UK
Utrecht University, The Netherlands

CAPRI and CASP

CAPRI

Since 2001

Critical Assessment of PRedicted **I**nteractions

CASP

Since 1994

Critical Assessment of Structure **P**redictions

Joint prediction rounds since 2014:

25 Targets	Round 30	CASP11	2014
10 Targets	Round 37	CASP12	2016
21 Targets	Round 46	CASP13	2018
12 Targets	Round 50	CASP14	2020
37 Targets	Round 54	CASP15	2022
34 Targets	Round 57	CASP16	2024

Prediction rounds on a “rolling” basis

Fits with publication schedule

3 to 4 weeks per prediction round

Fixed assessor team, established metrics

Prediction season

Intense 2 to 3 months

Varying assessors, varying metrics

Difference in targets

Mostly hetero-dimers or -trimers

Peptides, sugars, water positions

Mostly obligate, many homo-oligomers

Very large assemblies

Incites method development

Large-scale testing of methodologies

Website

CAPRI

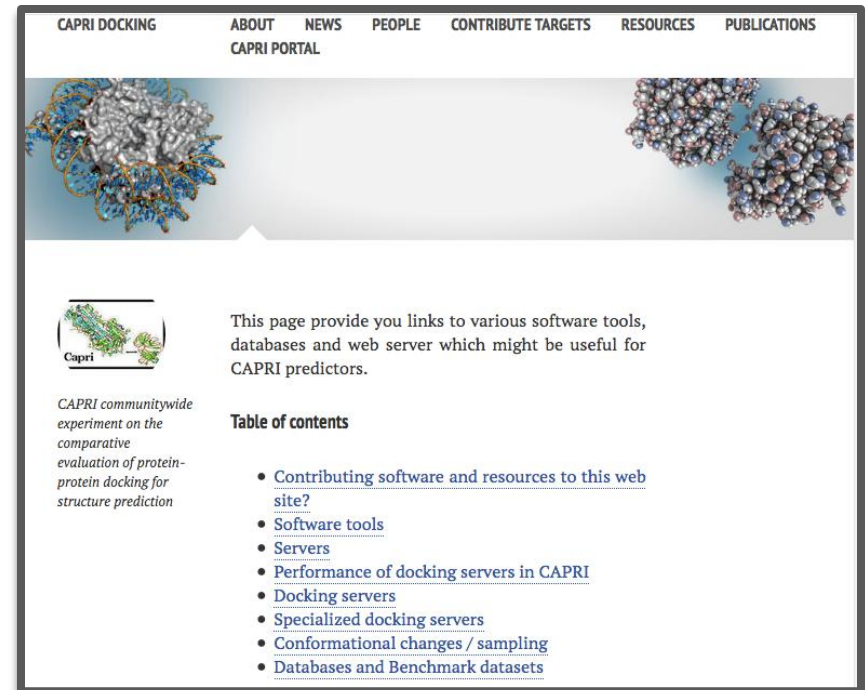
Since 2001

Critical **A**ssessment of
Predicted **I**nteractions

Protein Docking *and* Scoring

<https://www.pdbe.org/capri>
(for prediction submission)

<https://www.capri-docking.org/>
(community exchange portal)

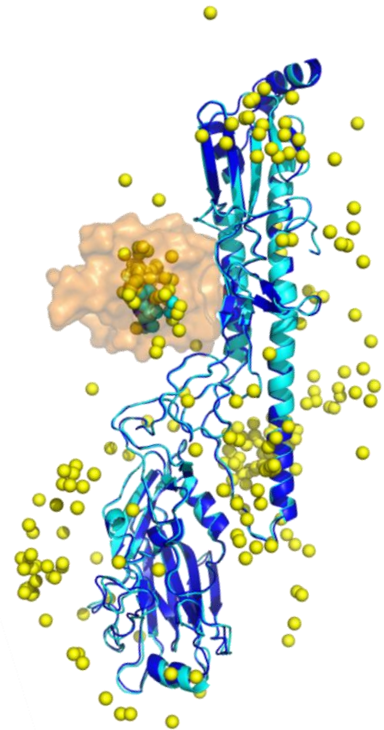


@CAPRIdock.bsky.social

CAPRI

Community-wide double blind experiment modelled after CASP, launched in 2001, aimed at assessing the performance of protein docking and scoring algorithms.

Prediction of the structure of an unpublished **protein-protein**, **protein-DNA/RNA**, **protein-peptide**, **protein-sugar** complex, or **multi-domain** protein; extended to the prediction of **binding affinity** and **interface water position**.



Marc



assessment

Alexandre



organization

Guillaume



website

Sameer

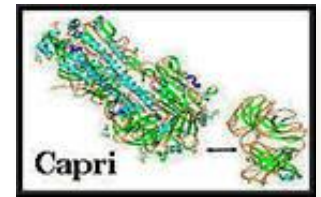


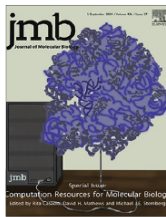
operations

EBI



infrastructure

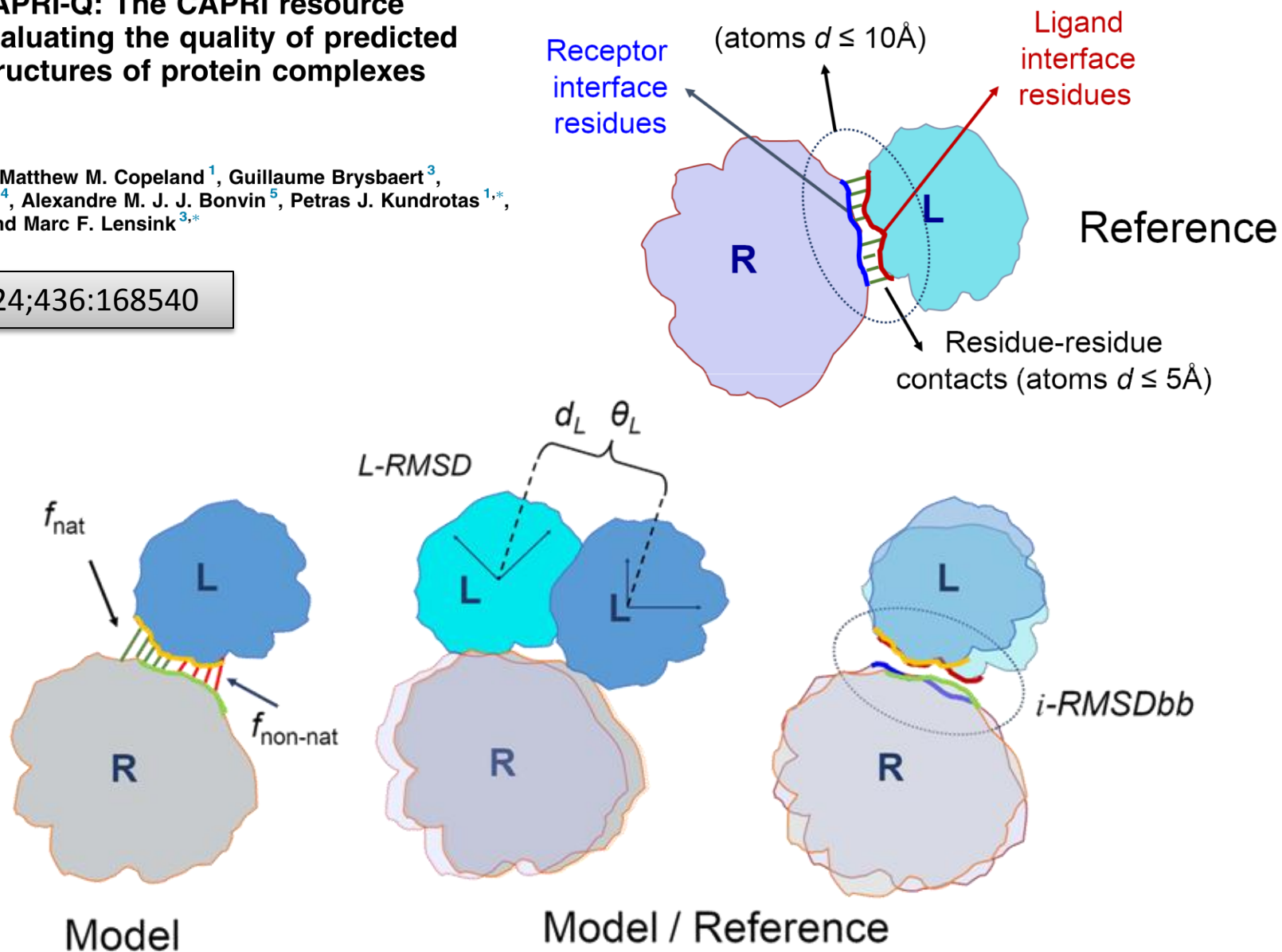




CAPRI-Q: The CAPRI resource evaluating the quality of predicted structures of protein complexes

Keeley W. Collins¹, Matthew M. Copeland¹, Guillaume Brysbaert³, Shoshana J. Wodak⁴, Alexandre M. J. J. Bonvin⁵, Petras J. Kundrotas^{1,*}, Ilya A. Vakser^{1,2,*} and Marc F. Lensink^{3,*}

J Mol Biol 2024;436:168540



CAPRI Assessment

1. CAPRI assessment is
 - a) **receptor/ligand** and
 - b) **interface** based
 - **L-rms**
 - **f_{nat} ; i-rms**
2. Four assessment categories
 - **incorrect**, **acceptable**, **medium**, **high**
3. For multimeric targets, **each interface is assessed separately**; depending on complexity, targets may then be split up into several assessment units (AU), with an AU representing a combination of individual interface scores
 - Either an AverageOf or BestOf
4. Final predictor score is the sum of these scores

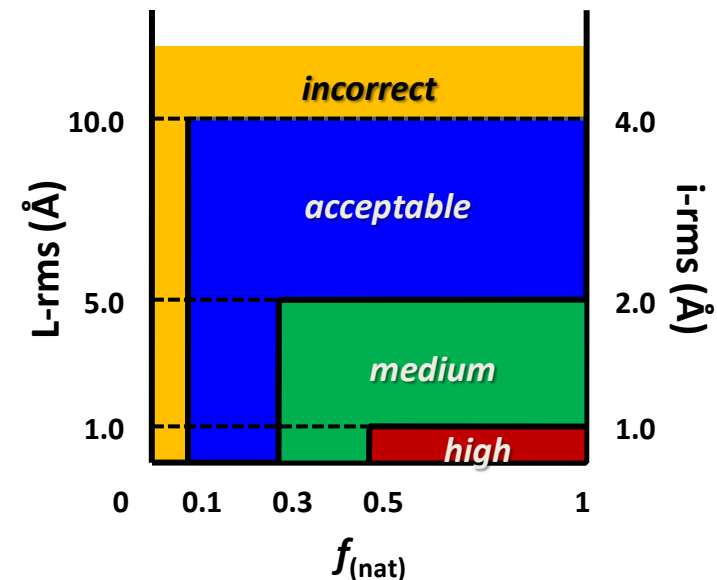
$$\text{Score} = \omega_1 \cdot N_{\text{ACC}} + \omega_2 \cdot N_{\text{MED}} + \omega_3 \cdot N_{\text{HIGH}}$$

$$\omega_1 = 1; \omega_2 = 2; \omega_3 = 3$$

*Only **L-rms**, **i-rms** and f_{nat} are used to classify protein-protein interaction models in CAPRI.*

*Additional quantities are being calculated, such as **S-rms**, which are useful quality measures for protein-peptide interaction models.*

An additional condition may be placed on f_{nonnat} values in the future.



Timeline and Conventions

CASP	CAPRI	Duration
Stage0		
Stage1	CAPRI R57	2 weeks
MassiveFold input		
	CAPRI Scoring	1 week
Stage2	CAPRI R58	1 week

Only *L-rms*, *i-rms* and f_{nat} are used to classify protein-protein interaction models in CAPRI.

The rms thresholds are modified for protein-peptide interaction; high-quality requires:

$$f_{(\text{nat})} \geq 0.8$$

$$\text{L-rms} \leq 1.0 \text{ \AA}$$

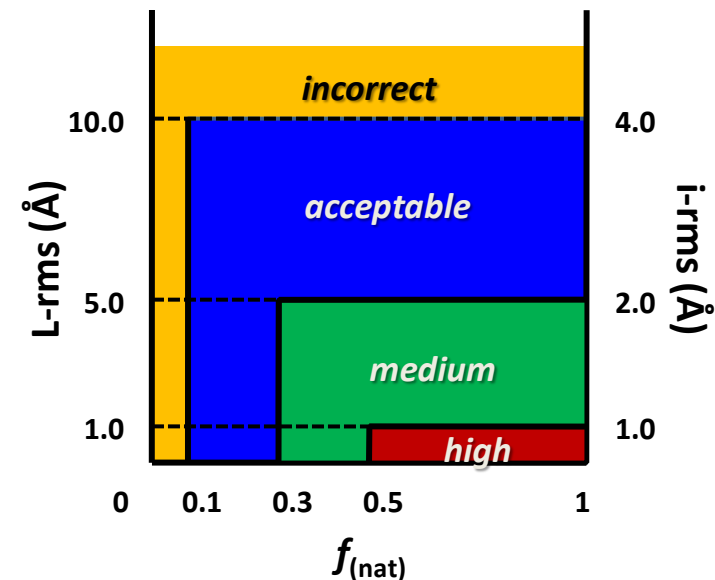
$$\text{i-rms} \leq 0.5 \text{ \AA}$$

1. Colors for the four assessment categories

- **Incorrect**
- **Acceptable**
- **Medium**
- **High**

2. Font style for the predictor groups

- Normal for human predictors
- **ALL-CAPS (and boldface) for servers**
- *S.Italics for scorer groups*
- Name⁺ means “and their server”

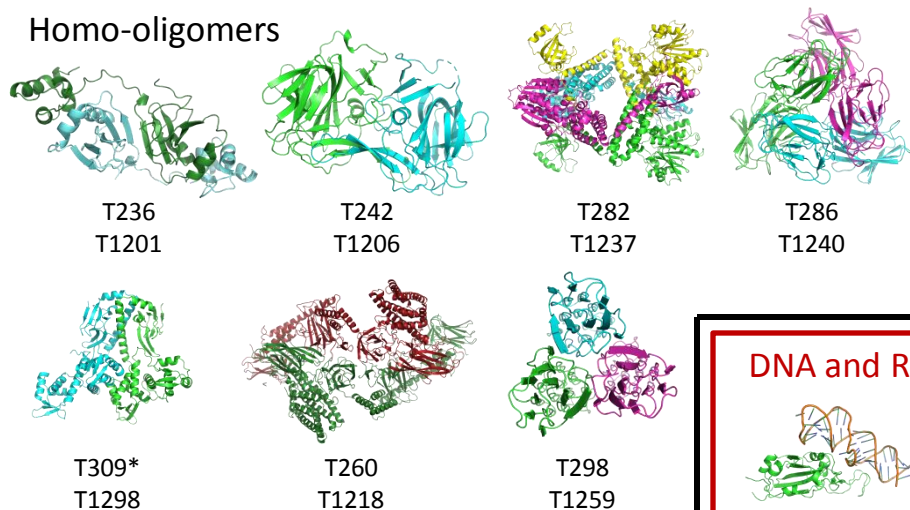


Statistics

CAPRI	T236 – T309		
CASP	T,H,M; 1201 – 2270		
Homo- and hetero-dimers	4	4	
Homo- and hetero-oligomers	4	15	
Antibodies and nucleic acids	10		
Number of targets	34		
Assessment units per stage	47	38	
Number of models (total)	88 198		
Number of models (top-5)	38 328		
MassiveFold models	430 486		
Total number of submitting groups	23	98	15
Submitting groups R57/stage1	15 – 22	23 – 77	11 – 13
Submitting groups R58/stage2	15 – 20	53 – 65	-
	CAPRI	CASP	Scorers

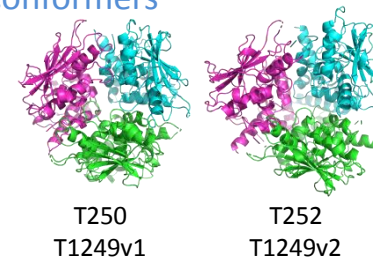
(included in CASP)

Homo-oligomers

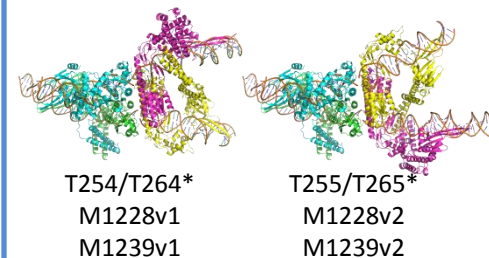
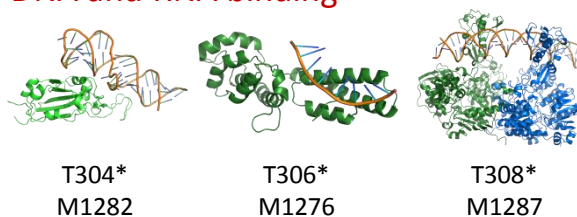


* No MassiveFold

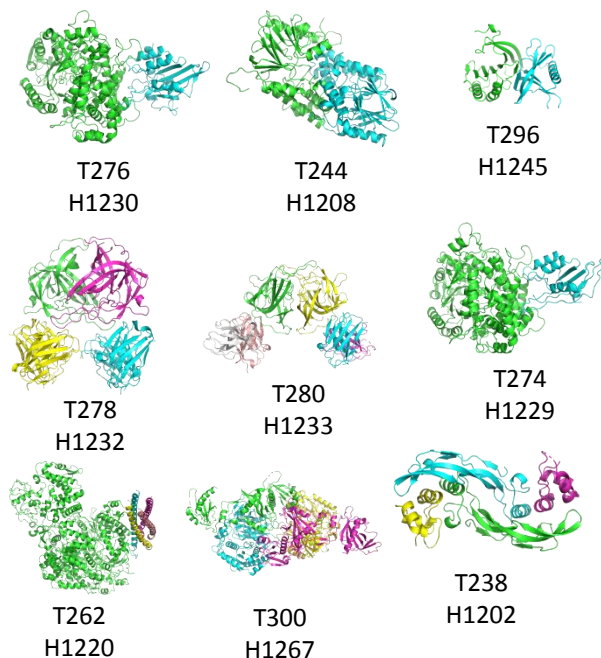
Conformers



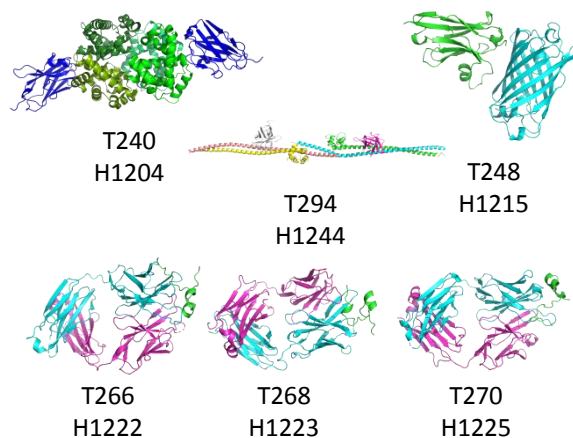
DNA and RNA binding



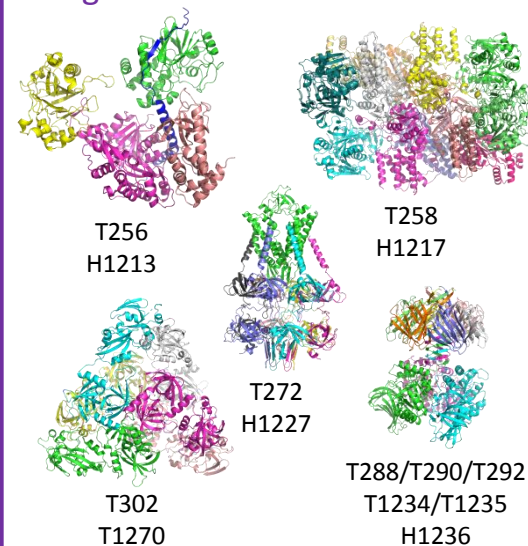
Hetero-oligomers



Antibodies and nanobodies



Large assemblies



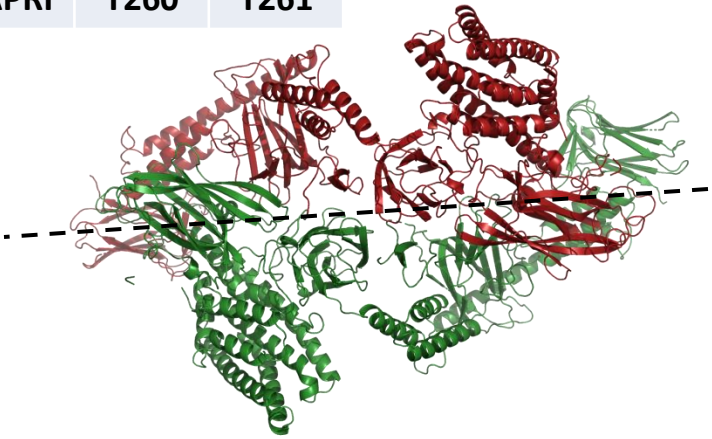
EXAMPLES

X-ray	2.9	A2
-------	-----	----

	Stage1	Stage2
CASP	T1218	T2218
CAPRI	T260	T261

#Contacts	Interface	Chains	Area
492	1	A:A'	2160

INT1 – “homo”, “dimer”



A difficult target
that was not so
difficult after all

DIFFICULT

AlphaFold ranked_0	
rms	51.05 Å
ipTM	0.515

Underlined and + means “and their servers”, i.e.

- Cheng

MULTICOM
- Kozakov

CLUSPRO
- Huang

HDOCK
- Kihara

LZERD
- Yang

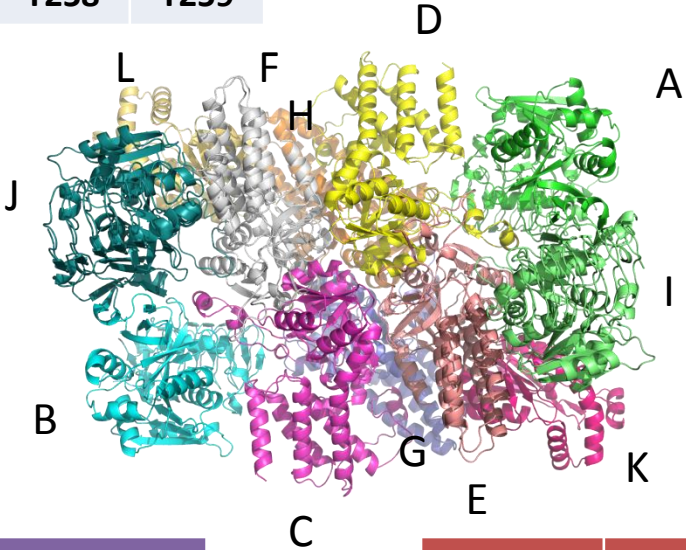
YANG-MULTIMER

Group stage1	Model	Fnat	L-rms	i-rms	Classification
<u>Yang</u> ⁺	1	0.68	3.53	2.17	Medium
Chang, <u>Cheng</u> ⁺ , <u>Kozakov</u> ⁺ , DEEPFOLD , PEZYFoldings, FTBiot0119					Medium
MassiveFold	First high quality model		Best quality model		Classification
0	-		-		Incorrect
Group stage2	Model	Fnat	L-rms	i-rms	Classification
<u>Yang</u> ⁺	5	0.68	3.53	2.17	Medium
<u>Huang</u> ⁺ , Chang, Kihara, <u>Kozakov</u> ⁺ , NKRNA-S , MIENSEMBLES, ZHENG , FTBiot0119, <u>Cheng</u> ⁺					Medium

EM	-	A2B2C2D2E2F2
----	---	--------------

	Stage1	Stage2
CASP	H1217	H2217
CAPRI	T258	T259

DIFFICULT



A=B
C=D
E=F
G=H
I=J
K=L

#Contacts	Interface	Chains	Area
227	1	C:F	2100
187	2 =	G:H	1750
186	3	F:J	1500
166	4	A:D	1250
155	5	C:G	1050
110	6	B:J	1000
99	7	H:L	720
113	8	C:E	700
107	9	E:K	600

average_of 1,2,3,4,5,6,7,8,9 – “large”

AlphaFold ranked_0	
rms	x Å
ipTM	0.663

Top-1

Interface	T258 CAPRI	T258 CASP	T258 Scorers	T259 CAPRI	T259 CASP	MassiveFold*	
1	20/19***/1**	66/64***/2**	12***	15***	57/56***/1**	146***	High
2	13/12***/1**	35/31***/4**	9***	14***	40/35***/5**	137***	High
3	16/12***/4**	58/39***/17**	12/5***/7**	14/9***/4**	53/41***/8**	52***	High
4	19/17***/2**	64/61***/2**	12/10***/2**	15***	56/55***	139***	High
5	20/15***/4**	65/46***/17**	12/9***/3**	15/8***/7**	57/32***/24**	98***	High
6	17/16***/1**	59/54***/5**	12***	14/13***	49/46***/4**	138***	High
7	1	7/2**	5/3**	5/3**	10/4**	0	Incorrect
8	15/10***/2**	50/29***/12**	10/4***/5**	15/9***/4**	49/32***/7**	92***	High
9	0	6/2**	0	1	8/3**	0	Incorrect

* Reduced set of 395 models

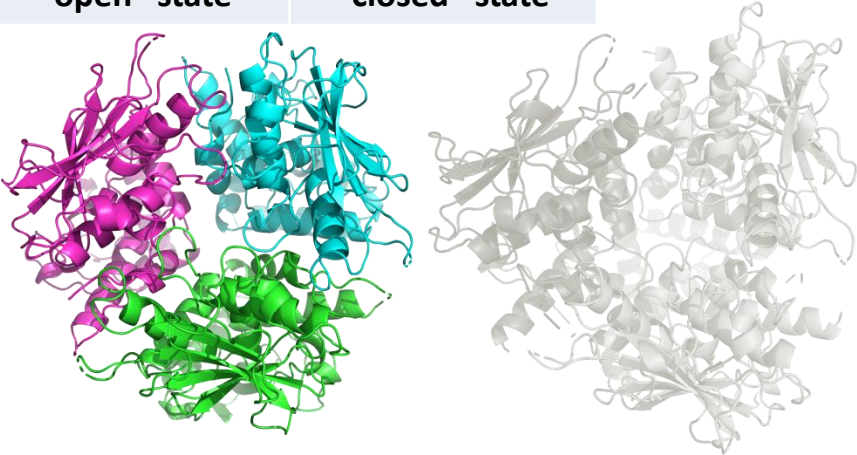
Another target
thought to be
difficult but
turned out easy

EM	-	A3
----	---	----

	Stage1	Stage2	Stage1	Stage2
CASP	T1249v1	T2249v1	T1249v2	T1249v2
CAPRI	T250	T251	T252	T253
	“open” state		“closed” state	

#Contacts	Interface	Chains	Area
513	T250	A:B	1110
517	T252	A:B	1040

INT1 – “homo”, “trimer”, “conformer”



“open” state

AlphaFold ranked_0	
rms	1.14 Å
ipTM	0.702

EASY

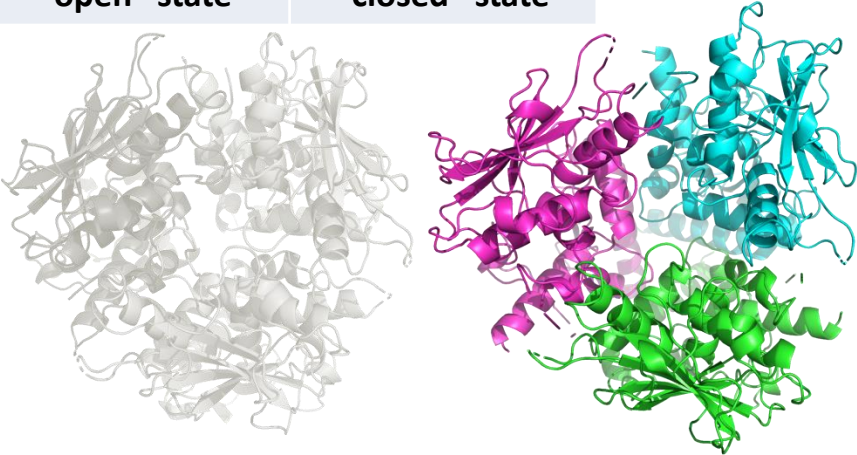


DIFFICULT

Group stage1			Classification
Giulini, Cool-PSP, LCDD-team, MRAFold, LupengKong, PEZYFoldings, Wallner, Bhattacharya, GHZ-Man, LZERD, YANG-MULTIMER , <i>Cheng, Giulini</i>			Medium
MassiveFold	Medium quality model	Best quality model	Classification
147/1**	#164	#164	Medium
Group stage2			Classification
Cool-PSP, LCDD-team, MRAFold, LupengKong, FTBiot0119, RAGfold-Prot1, ShanghaiTech-human, DEEPFOLD-SERVER, YANG-MULTIMER			Medium

EM	-	A3
----	---	----

	Stage1	Stage2	Stage1	Stage2
CASP	T1249v1	T2249v1	T1249v2	T1249v2
CAPRI	T250	T251	T252	T253
	“open” state		“closed” state	



AlphaFold ranked_0	
rms	1.14 Å
ipTM	0.702

EASY



DIFFICULT

#Contacts	Interface	Chains	Area
513	T250	A:B	1110
517	T252	A:B	1040

INT1 – “homo”, “trimer”, “conformer”

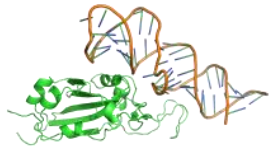
The “closed” state was difficult to predict, and not caught at all by the MassiveFold data

Giulini was the *only* predictor who found both states!

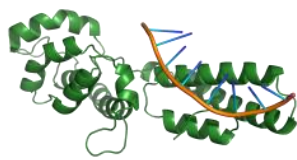
“closed” state

Group stage1			Classification
Vakser, Giulini, Huang, Pierce, plmfold, GuijunLab-QA, Yang-Server, Elofsson, MDOCKPP, HDOCK, MQA-SERVER, GHZ-ISM, UNICORN, PTQ			Acceptable
MassiveFold	Medium quality model	Best quality model	Classification
0	-	-	Incorrect
Group stage2			Classification
Giulini, Huang, Kihara, GuijunLab-QA, Yang-Server, Elofsson, MDOCKPP, HDOCK, UNICORN, GHZ-ISM, PTQ, MILLISECONDS			Acceptable

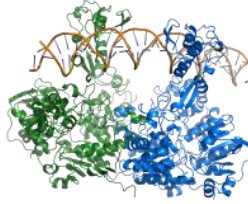
DNA and RNA binding



T304*
M1282



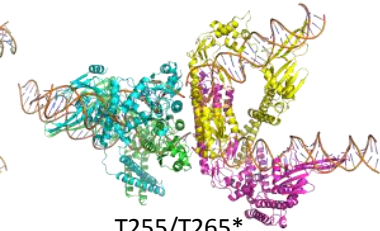
T306*
M1276



T308*
M1287



T254/T264*
M1228v1
M1239v1



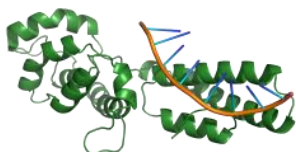
T255/T265*
M1228v2
M1239v2

DNA AND RNA BINDING

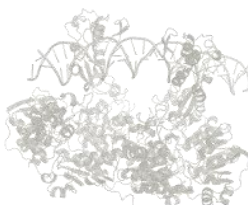
DNA and RNA binding



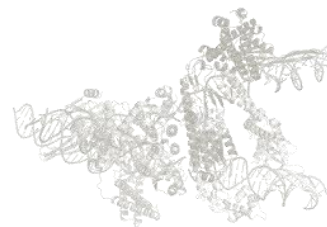
T304*
M1282



T306*
M1276



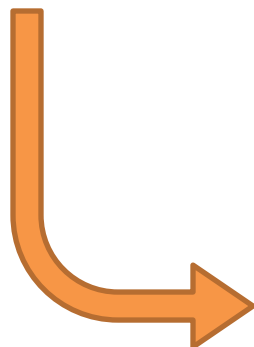
T308*
M1287



T254/T264*
M1228v1
M1239v1



T255/T265*
M1228v2
M1239v2



T304	M1282	No acceptable solutions	Highest DockQ	0.0544
T306	M1276	No acceptable solutions	Highest DockQ	0.0314

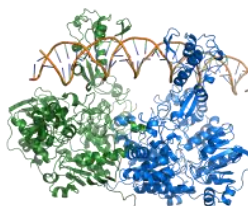
DNA and RNA binding



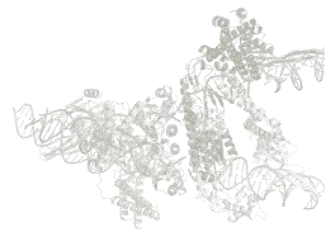
T304*
M1282



T306*
M1276



T308*
M1287



T254/T264*
M1228v1
M1239v1



T255/T265*
M1228v2
M1239v2

Predictors

Bhattacharya

Fernandez-Recio

ALPHAFOLD3-SERVER

B-LAB

GeneSilico

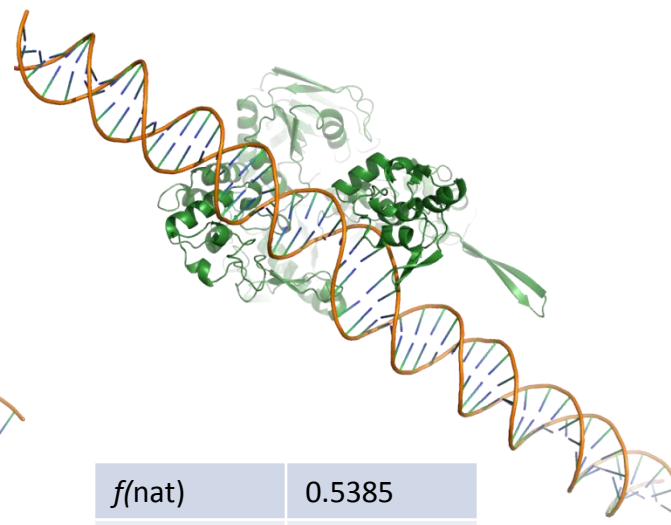
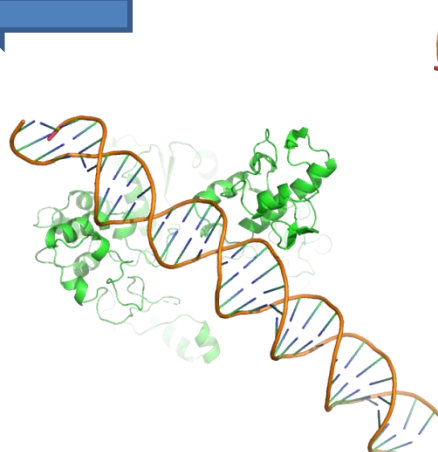
Diff

Scorers

Fernandez-Recio

Oliva

Kihara / LZERD



$f(\text{nat})$	0.5385
$f(\text{nonnat})$	0.8454
$i\text{-rms}$	3.8529

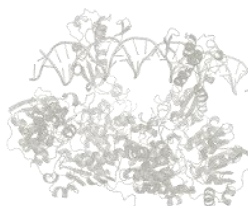
DNA and RNA binding



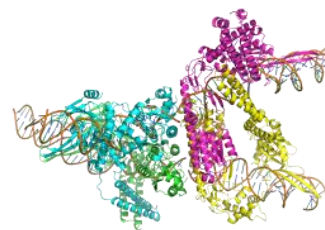
T304*
M1282



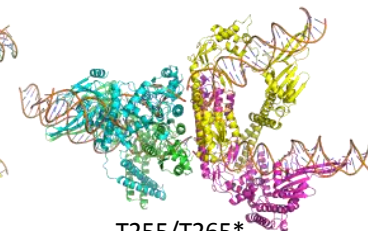
T306*
M1276



T308*
M1287



T254/T264*
M1228v1
M1239v1



T255/T265*
M1228v2
M1239v2

Predictors

CLUSPRO

Cool-PSP

Scorers

Zou

Oliva

Predictors

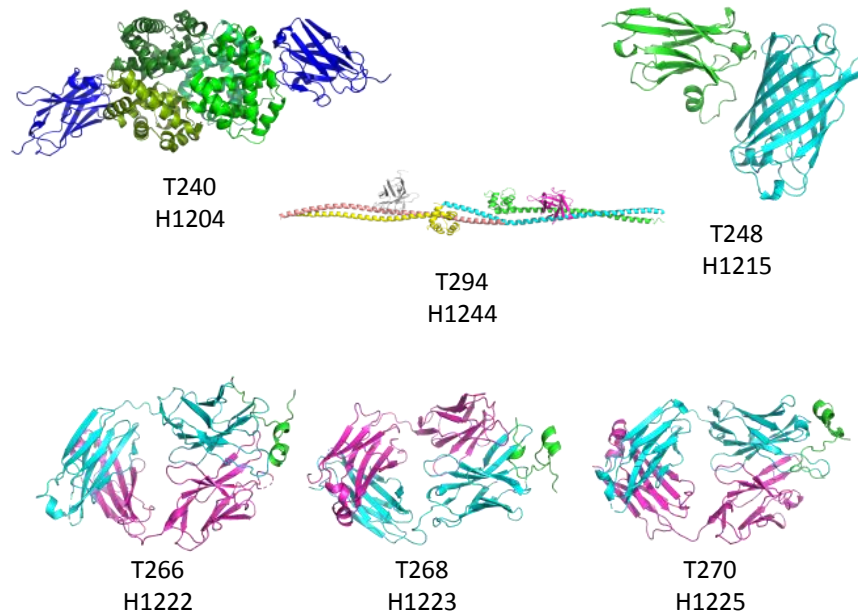
*Acceptable solutions
for **many** Predictors,
Scorers and Servers*

*The acceptable solutions are
mostly due to medium-quality
solutions for **T265***

Interface 1	Protein-DNA
Interface 2	Protein-DNA, alternate conformation
Interface 3	Protein-protein alternate conformation

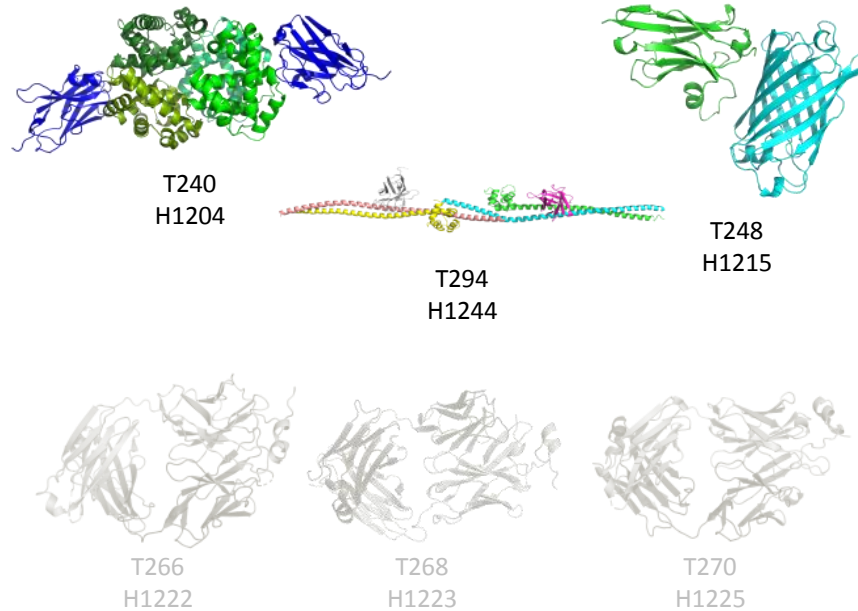
*No acceptable solutions for interface 1
Medium-quality for interfaces 2 and 3*

Antibodies and nanobodies



ANTIBODIES AND NANOBODIES

Antibodies and nanobodies



High

Kozakov

Acceptable

Cheng, Pierce, Zou

Fernandez-Recio

HYU-MLLab

LupengKong

MRAFold

MULTICOM (all variants)

OmniFold

YANG-MULTIMER, Yang, Yang-Server

MQA-SERVER

CSSB-Faker

FALCON2

Zou

High

Kozakov, **CLUSPRO**

Huang, **HDOCK**

Pierce, Brysbaert

Cheng, Cool-PSP

MENSEMBLES

Wallner, FTBiot0119

Zheng, **ZHENG-MM**, **NKRNA-S**

MULTICOM (all variants)

*Chang, Huang, **HDOCK**, **MDOCKPP***

Fernandez-Recio, Zou, Kihara

OpenComplex, elofsson, **OPENCOMPLEX**

ALPHAFOLD3-SERVER, CSSB-experimental

Zou, Kihara

No acceptable
solutions for
T294/H1244

X-ray	-	AB
-------	---	----

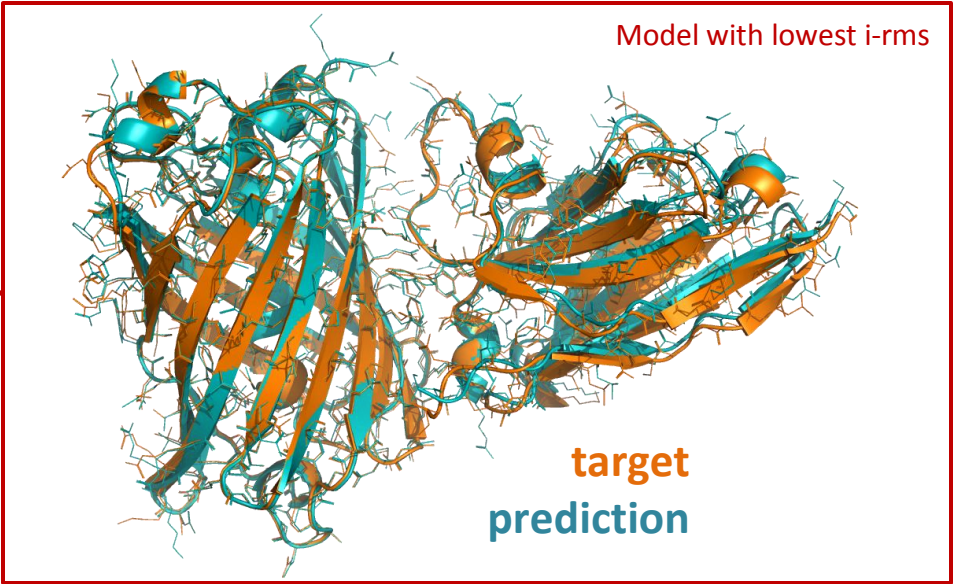
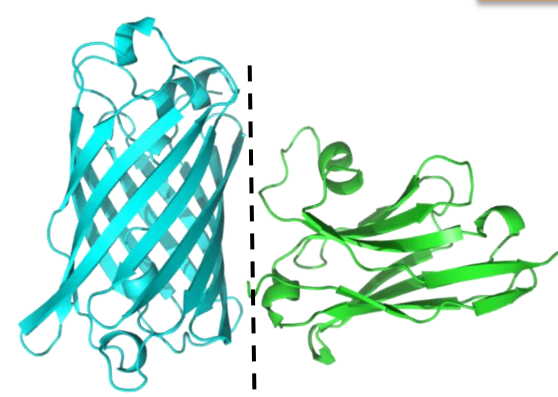
	Stage1	Stage2
CASP	H1215	T2215
CAPRI	T248	T249

#Contacts	Interface	Chains	Area
97	1	A:B	750

INT1 – “nanobody”

DIFFICULT

A target thought to be difficult but turned out easy



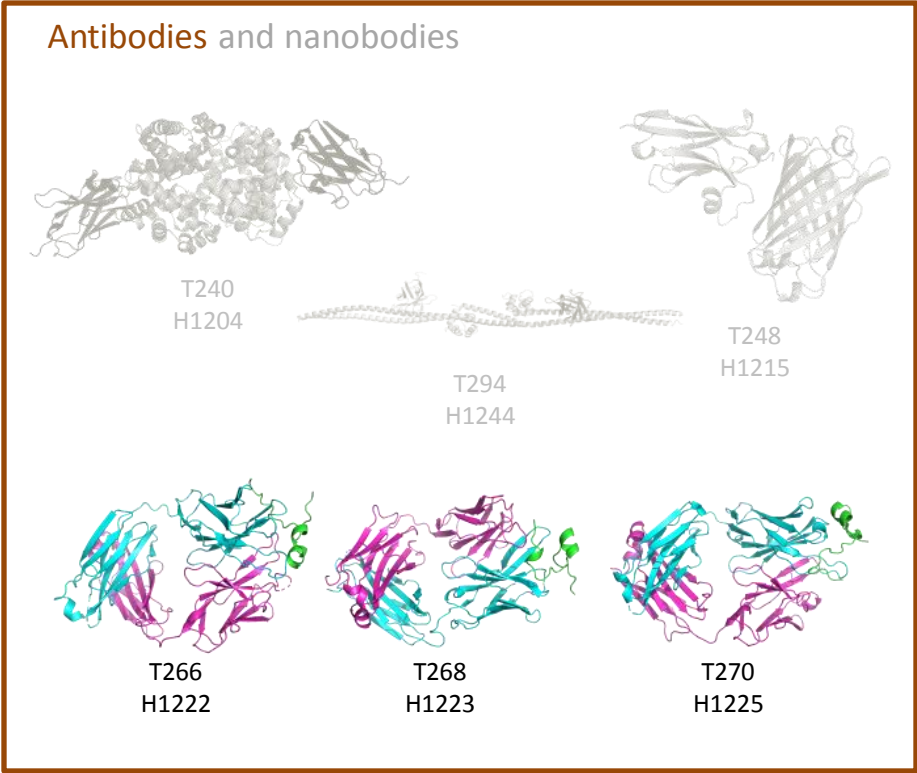
AlphaFold ranked_0	
rms	14.31 Å
ipTM	0.799

Underlined and + means “and their servers”, i.e.

- Cheng MULTICOM
- Kozakov CLUSPRO
- Huang HDOCK
- Kihara LZERD
- Zheng ZHENG-MULTIMER
- Yang YANG-MULTIMER

Group stage1	Model	Fnat	L-rms	i-rms	Classification
Brysbaert	5	0.90	1.71	0.35	High
Pierce, <u>Cheng</u> ⁺ , <u>Huang</u> ⁺ , <u>Kozakov</u> ⁺ , <u>Zheng</u> ⁺ , Wallner, +others, +some but not all Scorers					High
MassiveFold	First high quality model		Best quality model		Classification
204/29***/16**	#2		#5		High
Group stage2	Model	Fnat	L-rms	i-rms	Classification
Bates	4	0.86	1.43	0.33	High
Bates, Fernandez-Recio, <u>Kihara</u> ⁺ , <u>Kozakov</u> ⁺ , Chang, Zheng, <u>Yang</u> ⁺ , +others					High

Medium
Chang, Cheng, Gray, Kihara
Zou, MDOCKPP , Yang
Huang, HDOCK , Czaplewski
Kozakov, CLUSPRO , Elofsson
ALPHAFOLD3-SERVER
Chattacharya, PEZYFoldings
CSSB-Faker, CSSB-human
DEEPFOLD, DeepFold-refine
FALCON2, GHZ-ISM
GUIJNLAB (all variants)
LCDD-team, LinYang
MILLISECONDS, pimfold
MQA-SERVER, McGuffin
MULTICOM (all variants)
PTQ, UNICORN, smg-laval
XGroup, XGROUP
YANG-MM , Yang-Server
Giulini, Kihara, LZERD
Olechnovic, Zou, Chang
Huang, HDOCK , Shirali
Fernandez-Recio



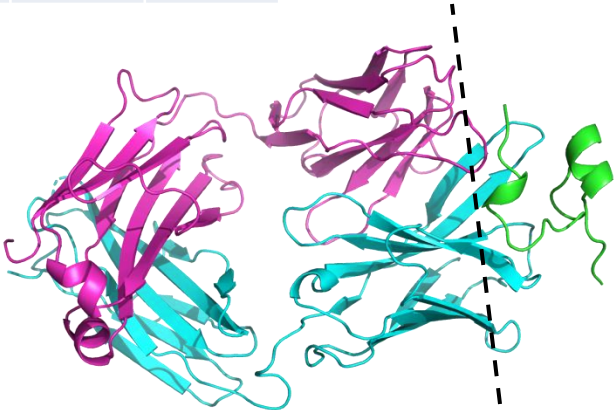
High
Kozakov, CLUSPRO , DeepFold-refine, DEEPFOLD-SERVER , LCDD-team, PEZYFoldings
Giulini, Olechnovic, LZERD
Acceptable
Huang, HDOCK , Chang, GromihaLab, OPENCOMPLEX , OpenComplex
Elofsson, RAGfold-Prot1, Bhattacharya, Mialab-prediction, FTBiot0119
Kihara, MDOCKPP

X-ray	-	A:HL
-------	---	------

	Stage1	Stage2
CASP	H1223	H2223
CAPRI	T268	T269

#Contacts	Interface	Chains	Area
341	1	A:HL	930

INT1 – “homo”, “dimer”



DIFFICULT

AlphaFold ranked_0	
rms	x Å
ipTM	crashed



Model with lowest i-rms

Group stage1	Model	Fnat	L-rms	i-rms	Classification
CLUSPRO	1	0.90	13.03	0.56	High
Kozakov, LCDD-team, <u>DeepFold</u> ⁺ , PEZYFoldings, <i>Olechnovic</i> , <i>Giulini</i> , LZERD					High
MassiveFold	Medium quality model		Best quality model		Classification
108/1**	#7707		#7707		medium
Group stage2	Model	Fnat	L-rms	i-rms	Classification
LZERD	2	0.92	0.88	0.85	High
<u>Cheng</u> ⁺ , Kihara, <u>Kozakov</u> ⁺ , LCDD-team, <u>DeepFold</u> ⁺ , PEZYFoldings, Wallner					High

Underlined and + means “and their servers”, i.e.

Cheng

MULTICOM

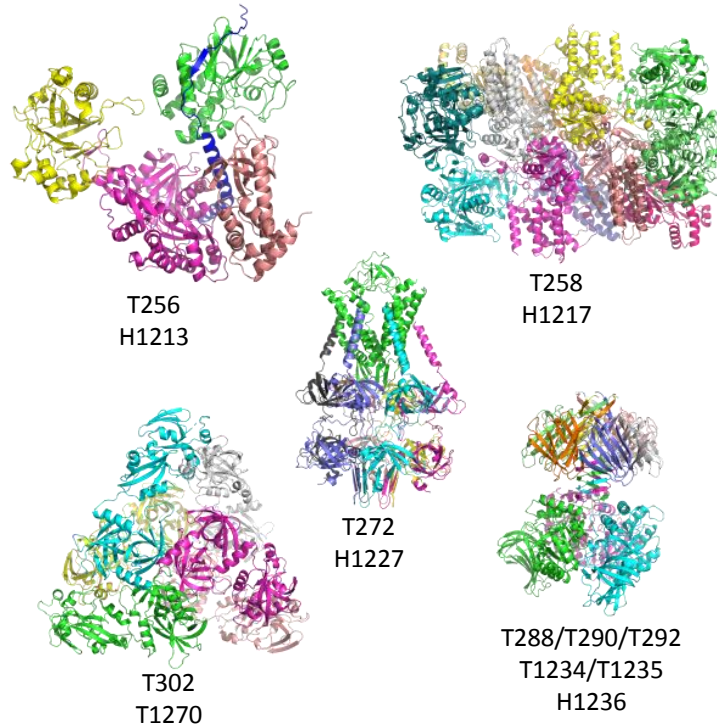
Kozakov

CLUSPRO

DeepFold-refine

DEEPFOLD

Large assemblies



LARGE ASSEMBLIES

Medium quality for many predictors

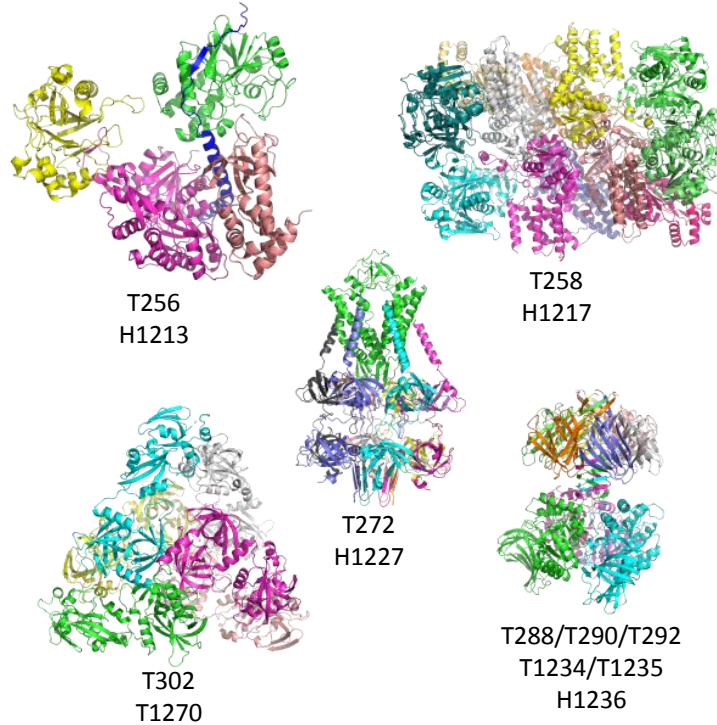
CSSB-experimental

Chang, Fernandez-Recio, Giulini, HYU-MLLab, **YANG-MM**, **MILLISECONDS**, GromihaLab, **MULTICOM**-{LLM,GATE}, Wallner, **OPENCOMPLEX**
Giulini, Fernandez-Recio

High quality for the primary interfaces, no acceptable solutions for the weak interface

The large majority of groups

Large assemblies



High quality for some predictors, medium for most

Cheng, Giulini, **MDOCKPP**, GromihaLab, **COLABFOLD**, **MULTICOM** (all variants), Bhattacharya, **MULTIFOLD2**

Medium quality for many predictors

Gray, Vakser

Kihara, Chang, Cheng, Czaplewski, Pierce, Fernandez-Recio, Huang, **HDOCK**, Zou, **MDOCKPP**, Schneidman, Brysbaert

Acceptable to medium quality for most predictors

Brysbaert, Fernandez-Recio

Kihara, **LZERD**, Zou, Chang, plmfold, MRAfold, LupengKong
Giulini, Zou, Kihara

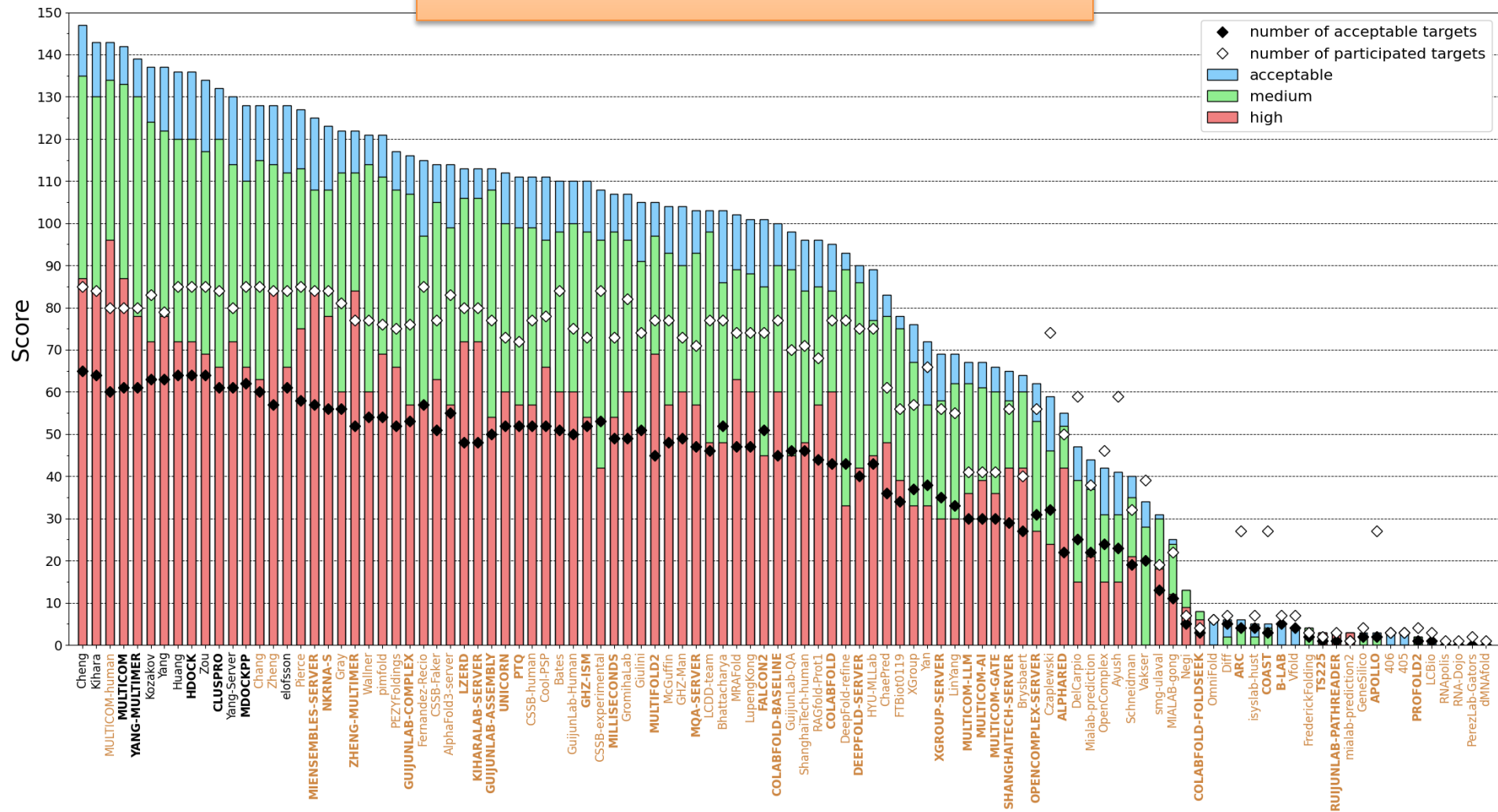
$$\text{Score} = \omega_1 \cdot N_{\text{ACC}} + \omega_2 \cdot N_{\text{MED}} + \omega_3 \cdot N_{\text{HIGH}}$$

$$\omega_1 = 1; \omega_2 = 2; \omega_3 = 3$$

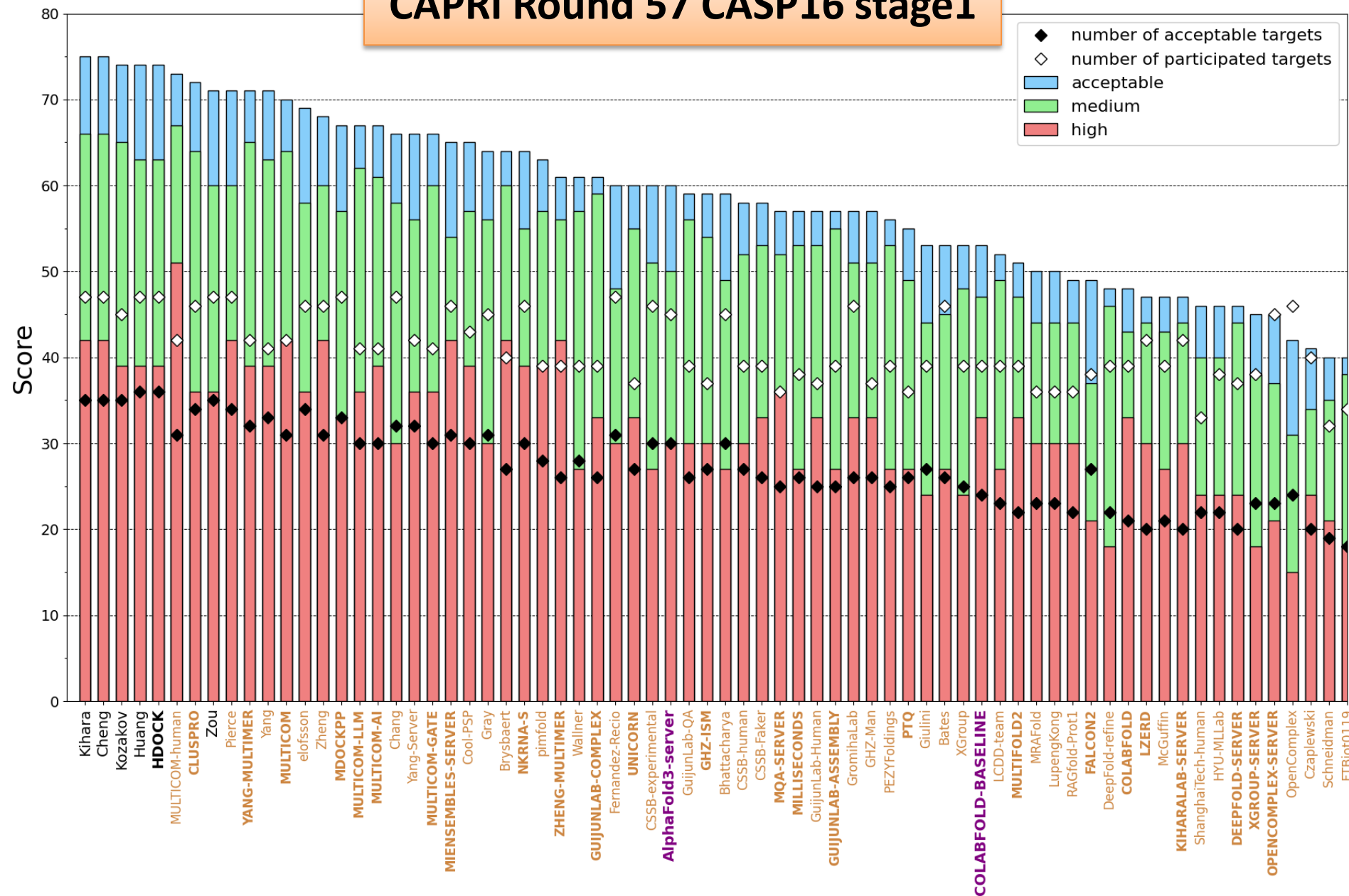
RANKING

CAPRI Rounds 57 & 58

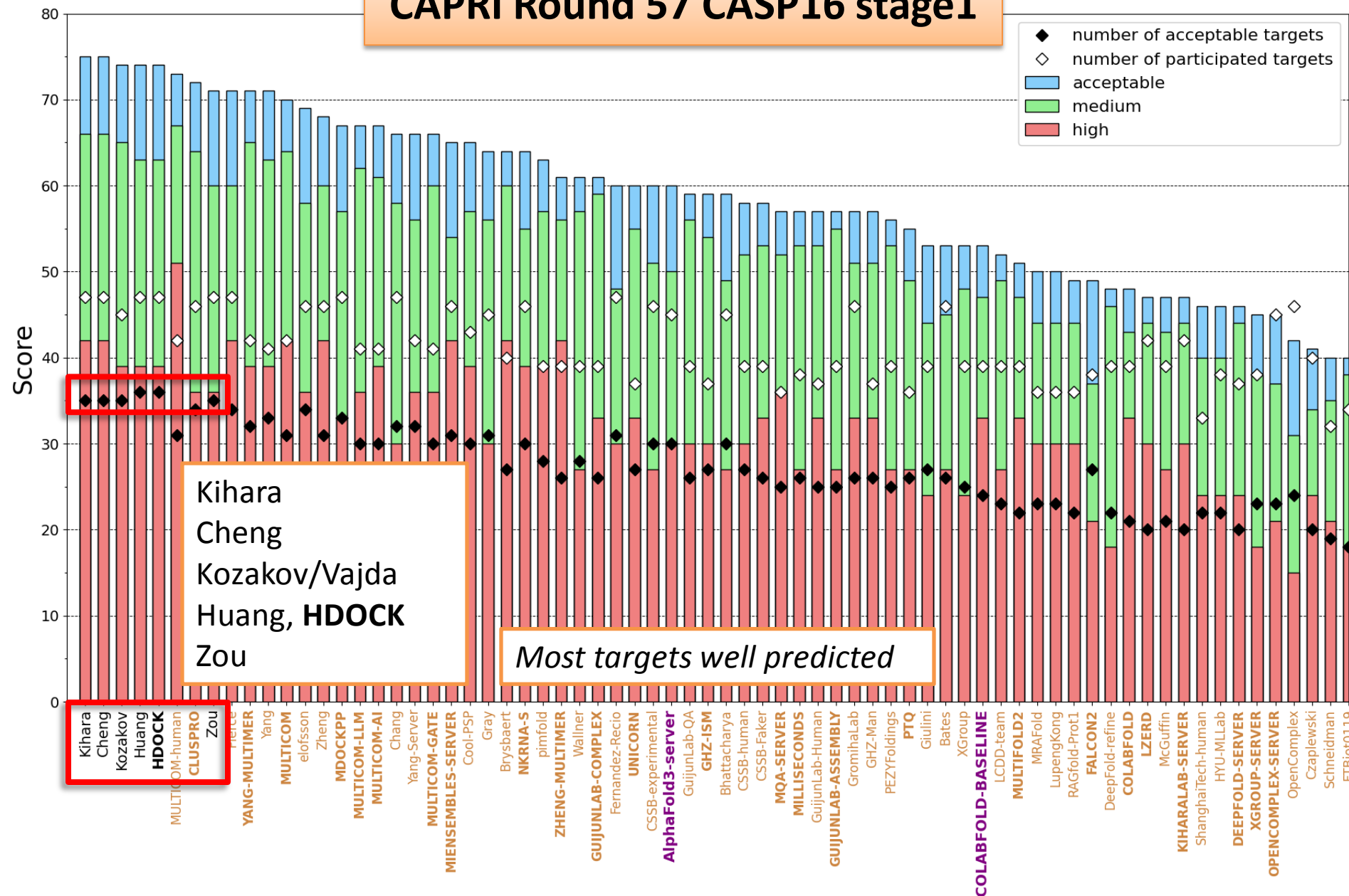
CASP16 stage1 & stage2



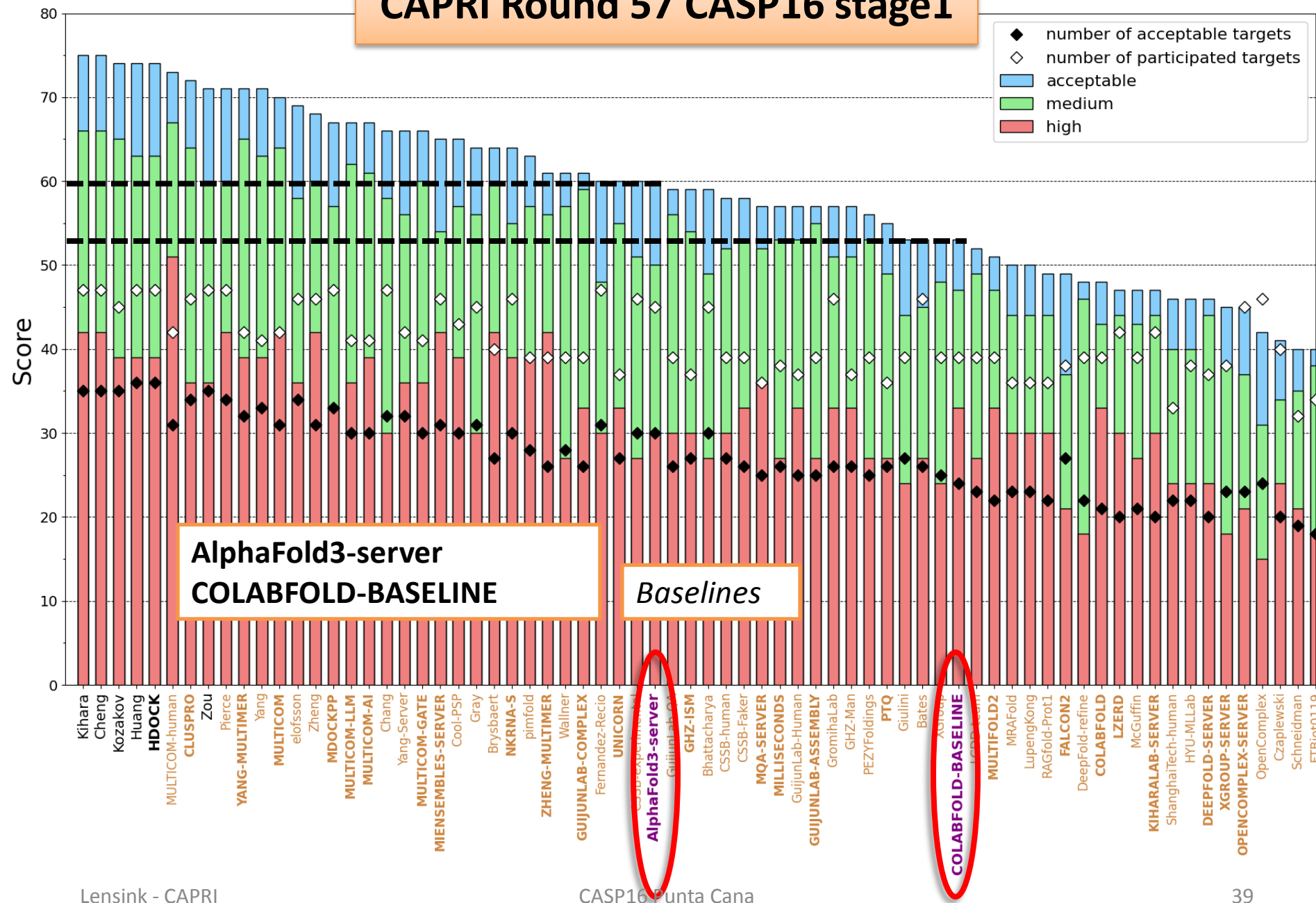
CAPRI Round 57 CASP16 stage1



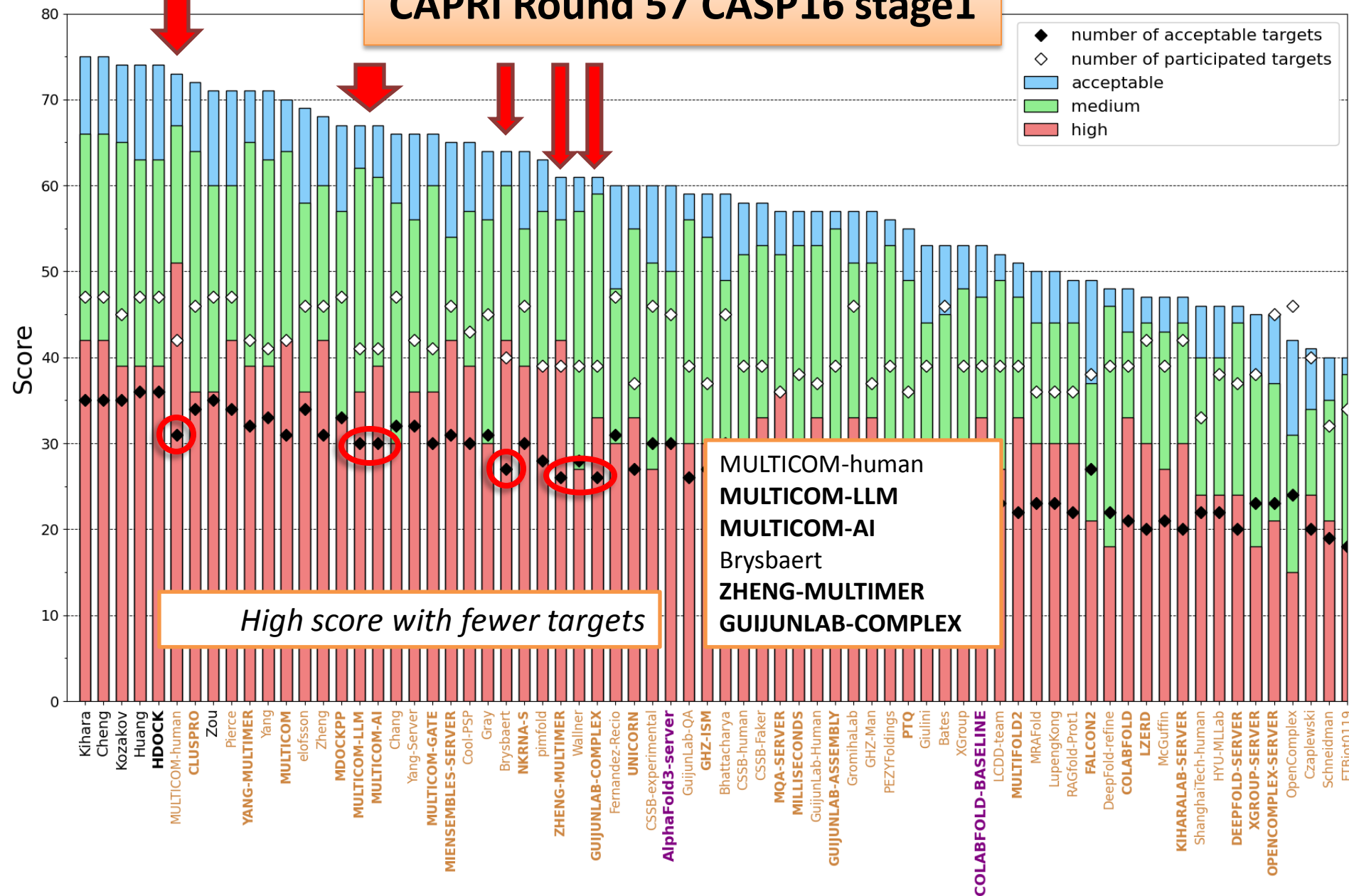
CAPRI Round 57 CASP16 stage1



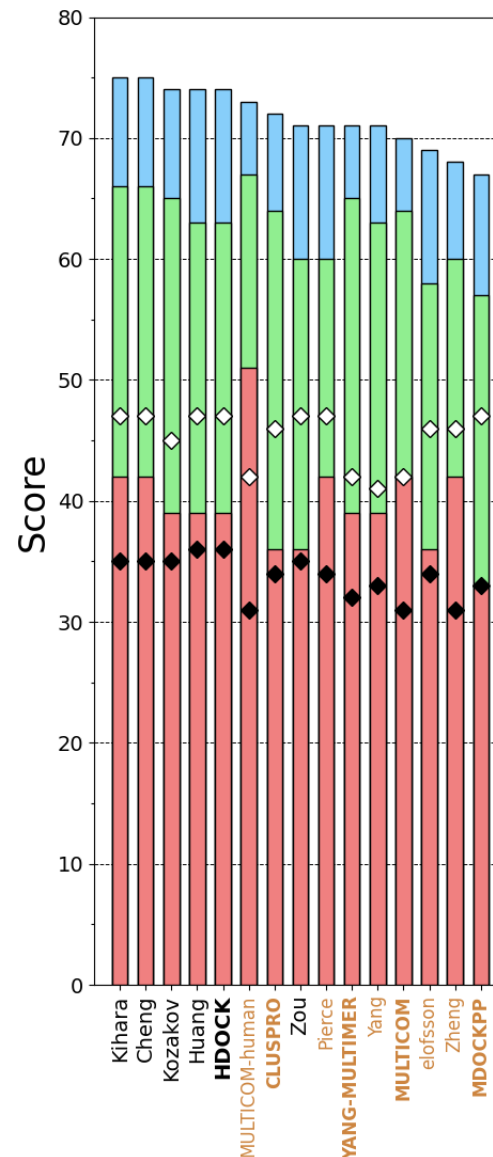
CAPRI Round 57 CASP16 stage1



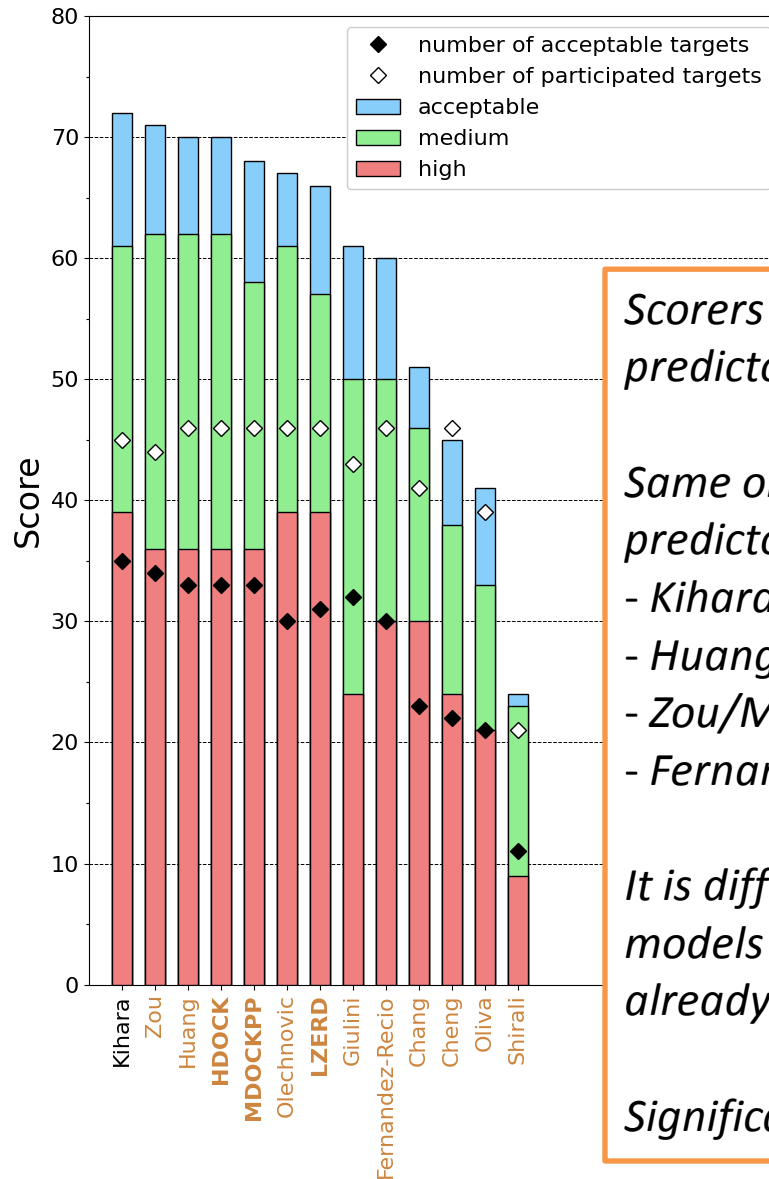
CAPRI Round 57 CASP16 stage1



Predictors



Scorers



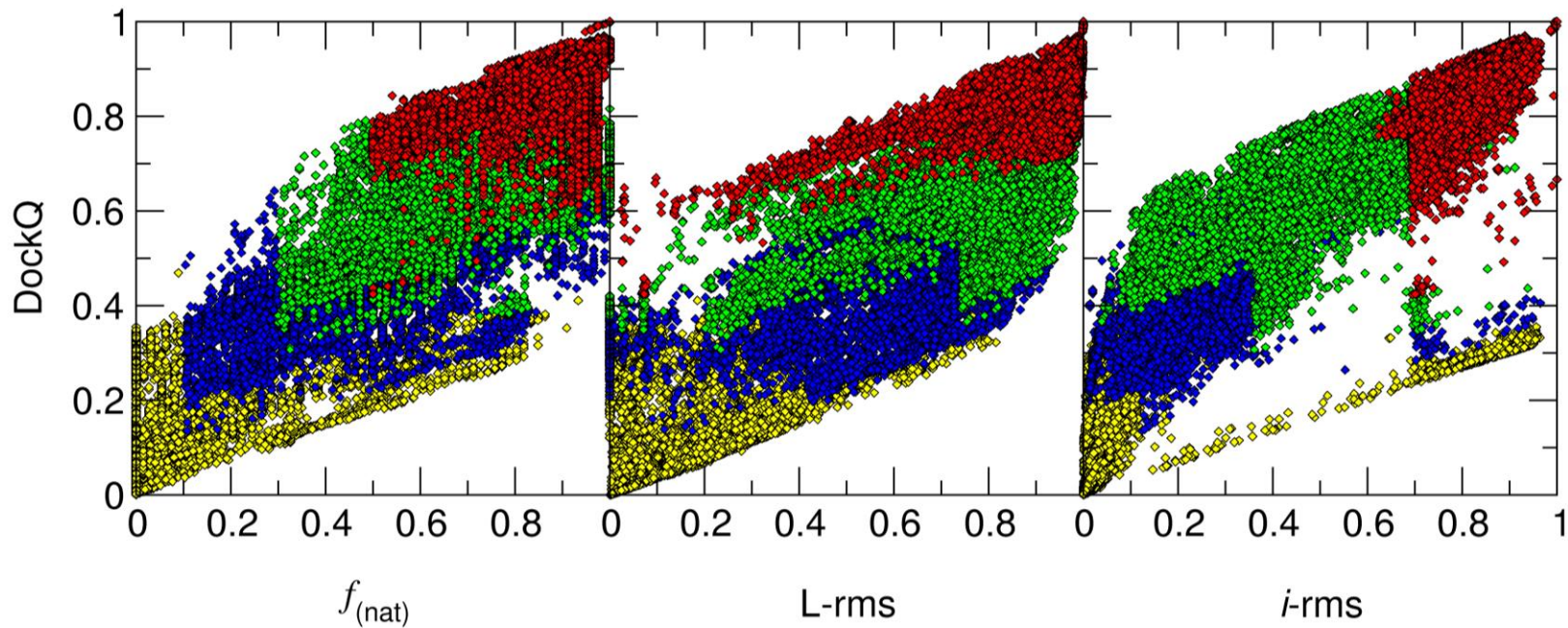
Scorers do not do a better job than predictors

Same or similar scores for predictor/scorer teams:

- Kihara
- Huang/HDOCK
- Zou/MDOCKPP
- Fernandez-Recio

It is difficult to identify the best models when most other are already very good ones

*Significant improvement for **LZERD***



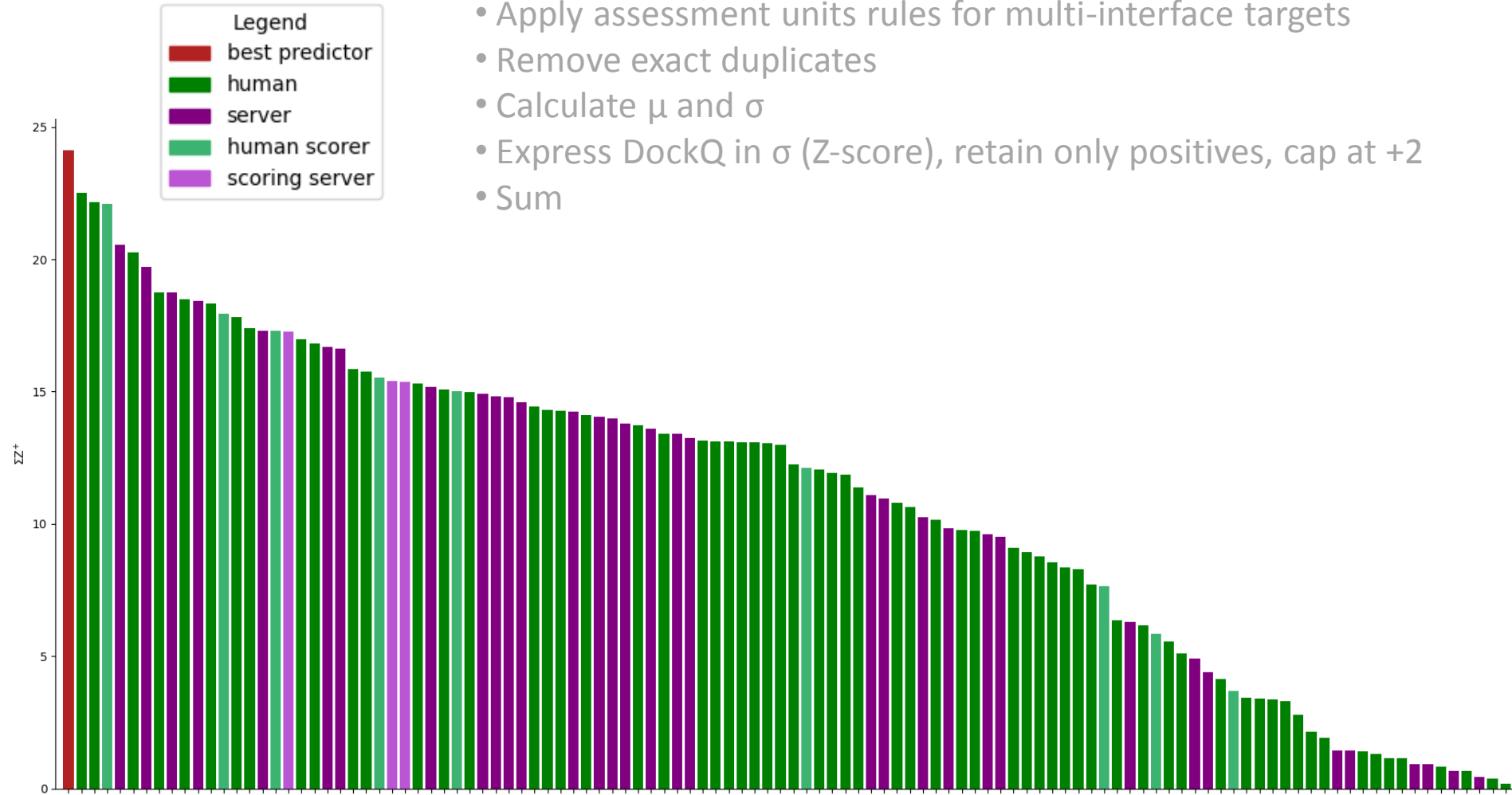
$$\text{DockQ} = \frac{1}{3} F(\text{nat}) + \frac{1}{3} \text{L-rms} + \frac{1}{3} \text{i-rms}$$

DOCKQ

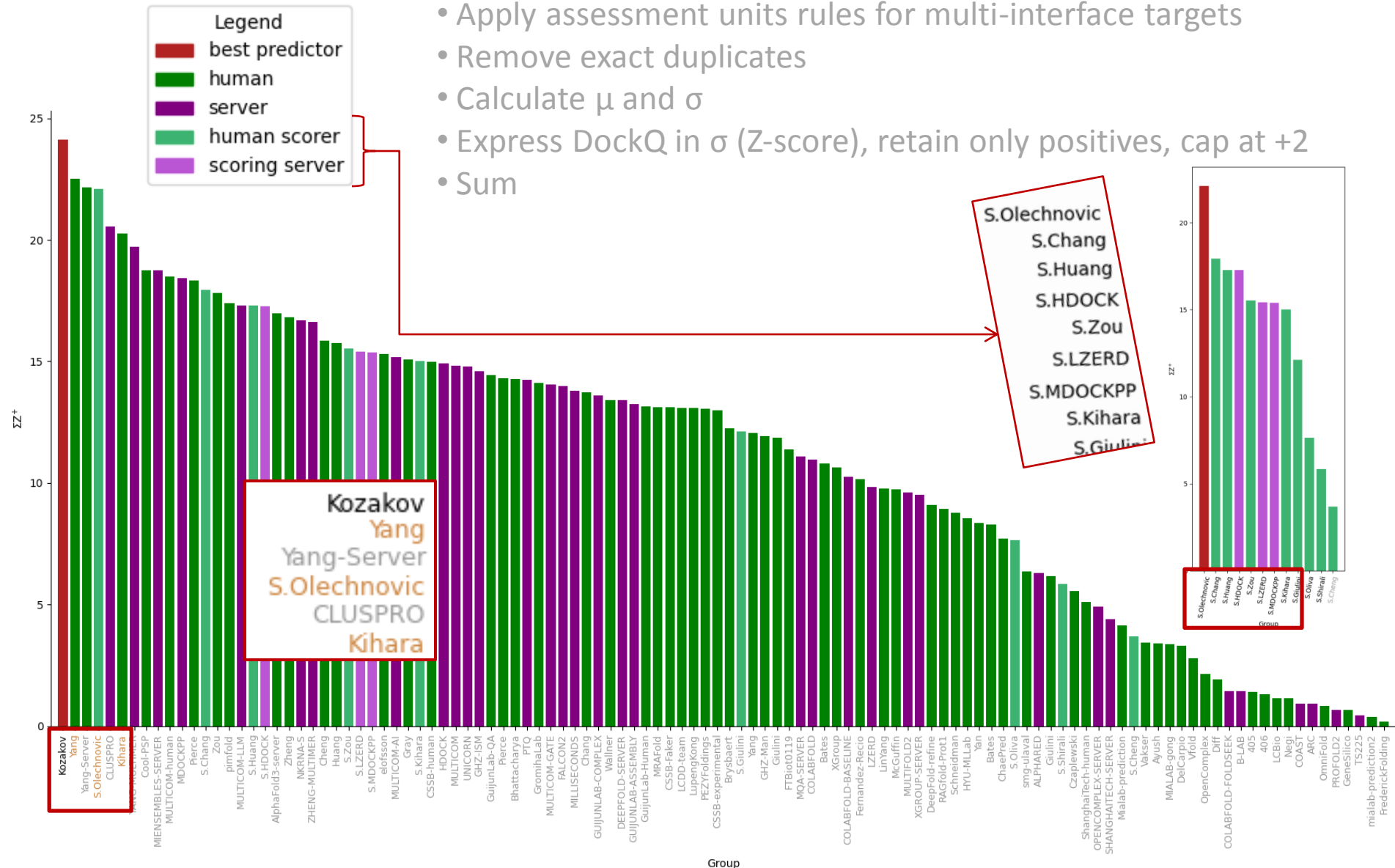
```
function rms_scaled(rms, d) {
  r = rms / d;
  r = 1.0 / (1.0 + r*r);
  return(r);
}
BEGIN { d1 = 8.5; d2 = 1.5; }
{ q = ($1 + rms_scaled($2, d1) + rms_scaled($3, d2)) / 3.0;
  printf "%.4f\n", q;
}
```

- Extract DockQ values for all participants' best top-5 model
- Apply assessment units rules for multi-interface targets
- Remove exact duplicates
- Calculate μ and σ
- Express DockQ in σ (Z-score), retain only positives, cap at +2
- Sum

- Extract DockQ values for all participants' best top-5 model
- Apply assessment units rules for multi-interface targets
- Remove exact duplicates
- Calculate μ and σ
- Express DockQ in σ (Z-score), retain only positives, cap at +2
- Sum

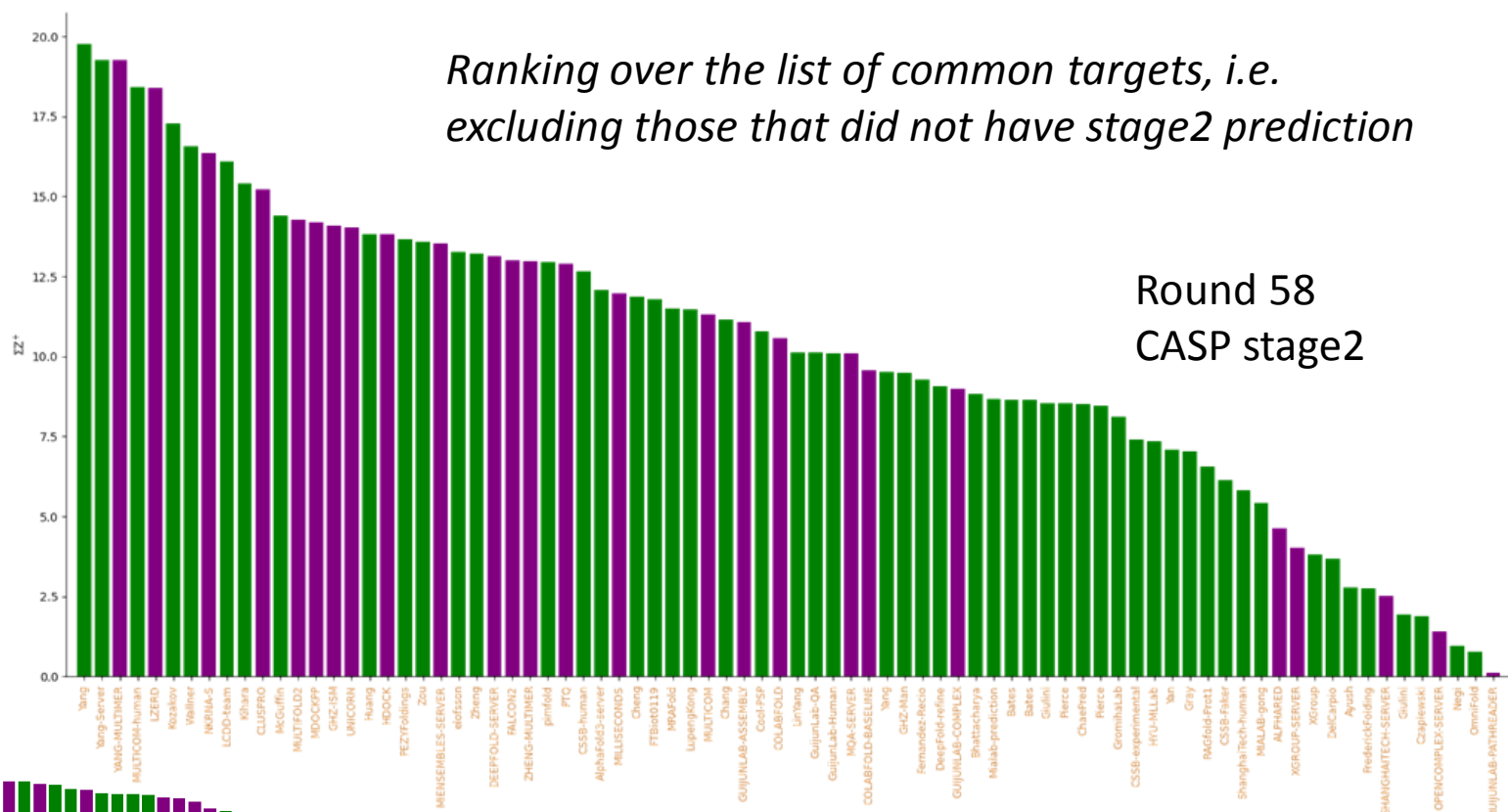


- Extract DockQ values for all participants' best top-5 model
- Apply assessment units rules for multi-interface targets
- Remove exact duplicates
- Calculate μ and σ
- Express DockQ in σ (Z-score), retain only positives, cap at +2
- Sum

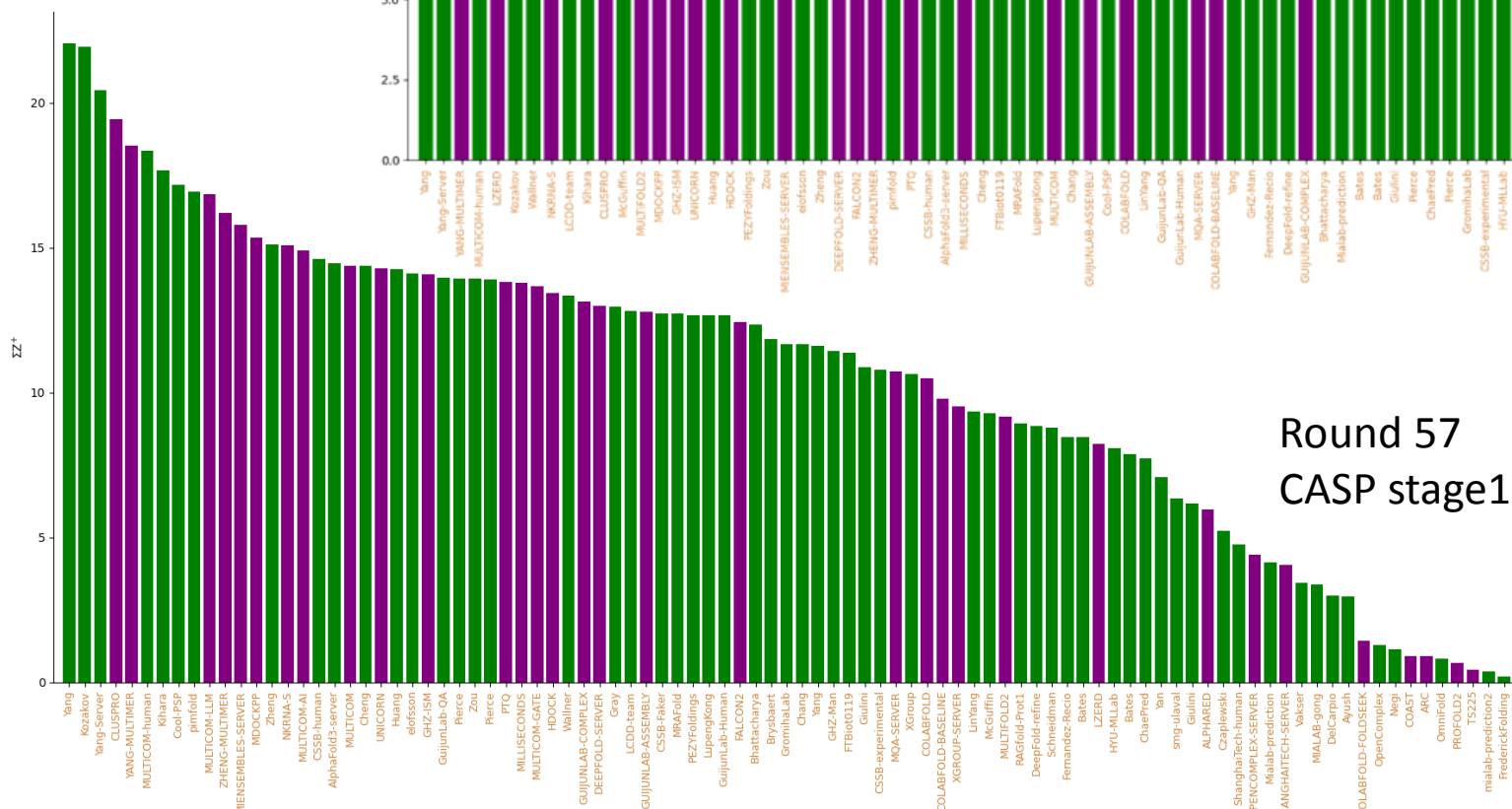


*Ranking over the list of common targets, i.e.
excluding those that did not have stage2 prediction*

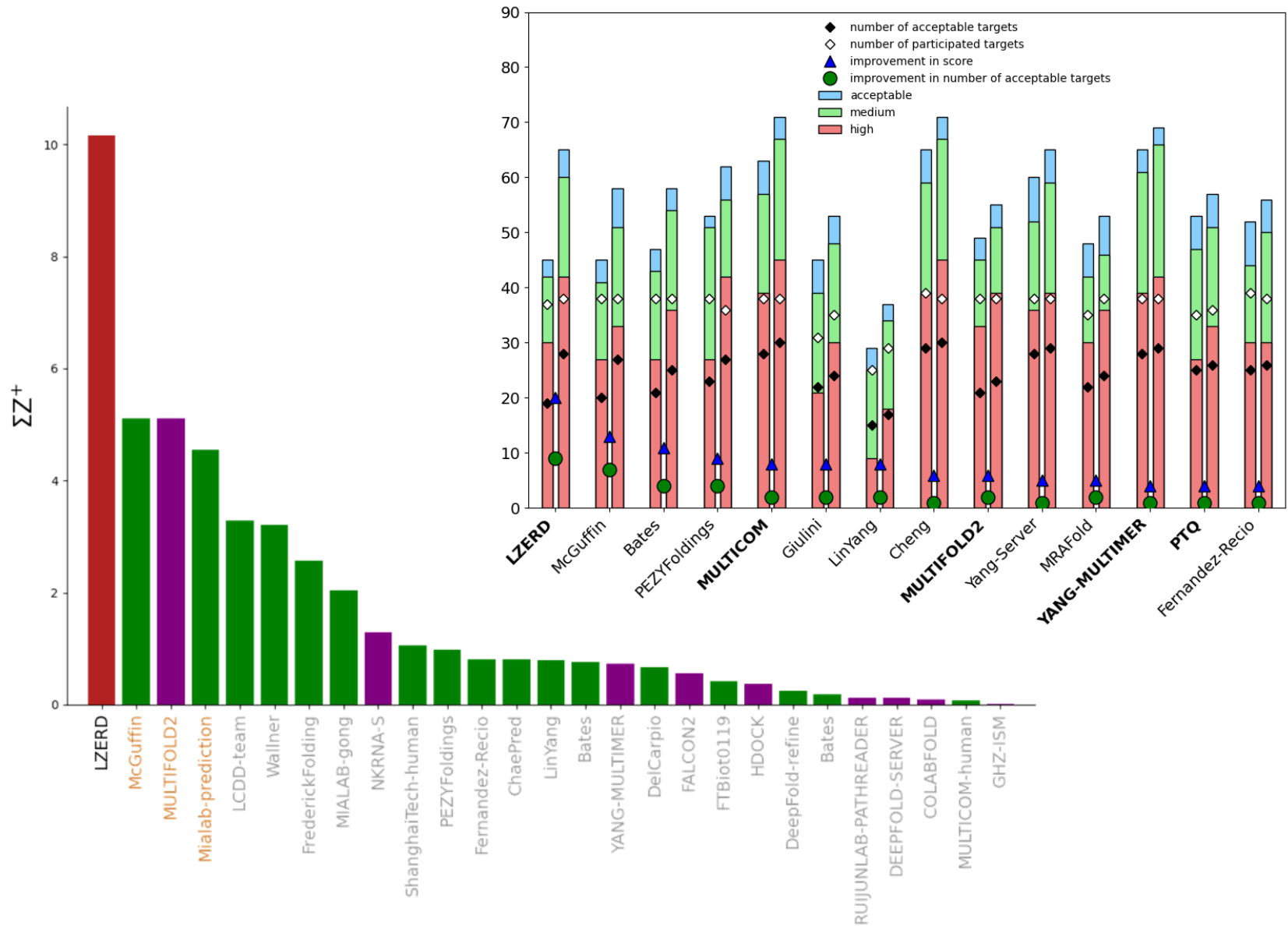
Round 58
CASP stage2



Round 57
CASP stage1

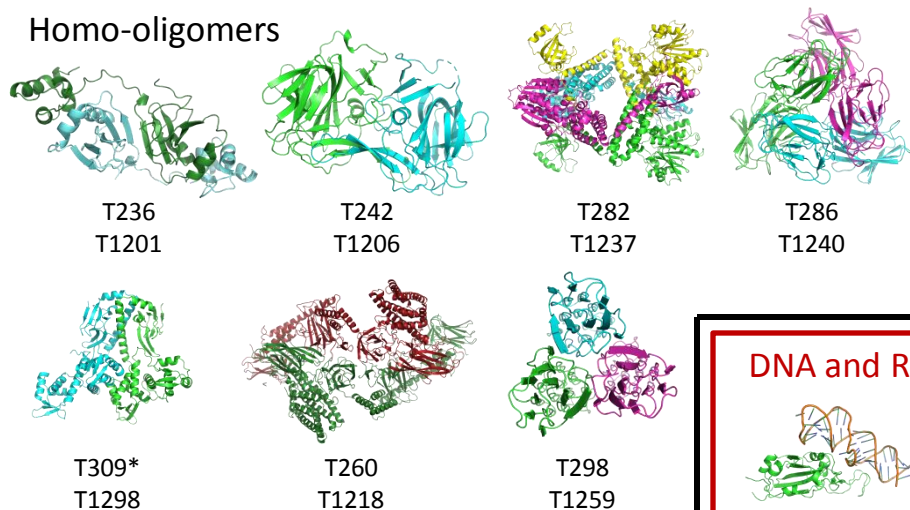


Ordered on improvement in score
 Minimum 20 targets participated
 Difference in participation R58 vs R57 no more than 4 targets



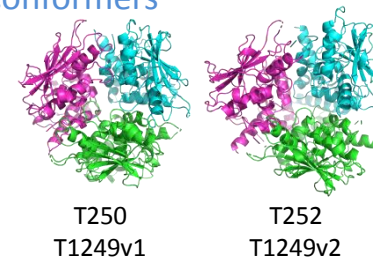
TARGETS THAT FAILED

Homo-oligomers

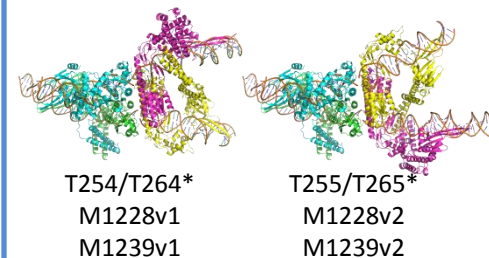
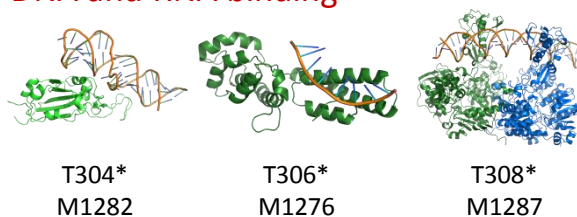


* No MassiveFold

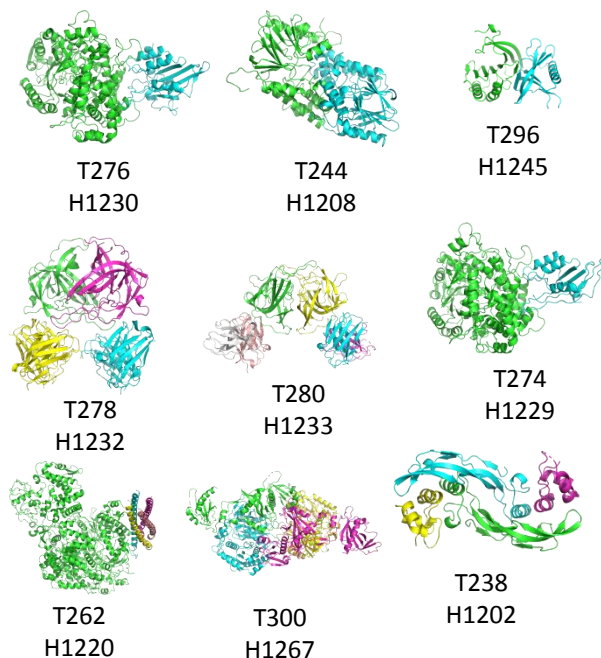
Conformers



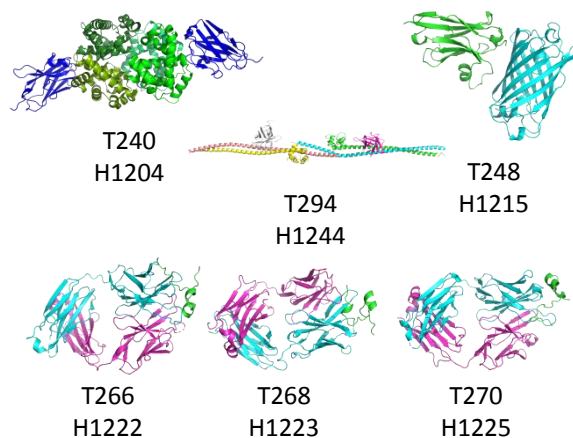
DNA and RNA binding



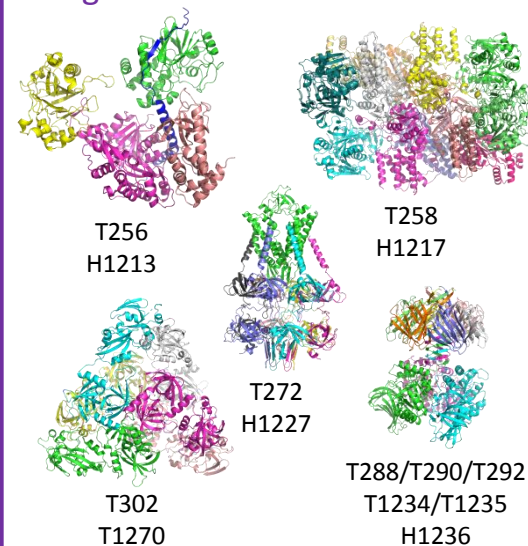
Hetero-oligomers



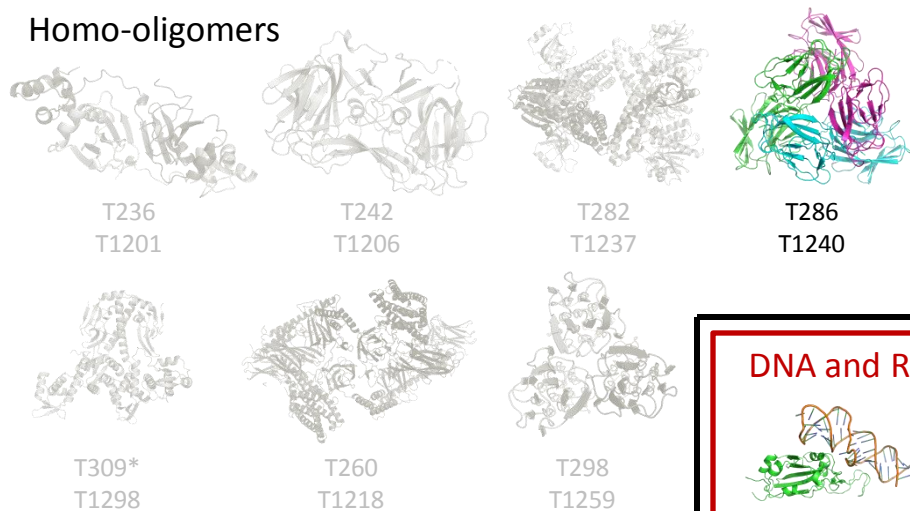
Antibodies and nanobodies



Large assemblies

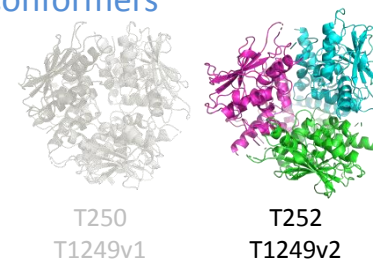


Homo-oligomers

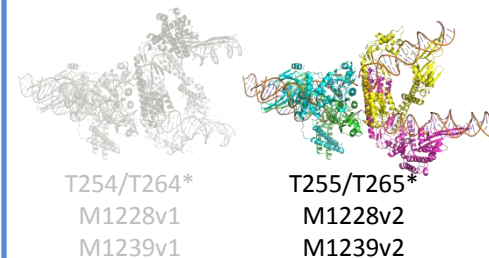
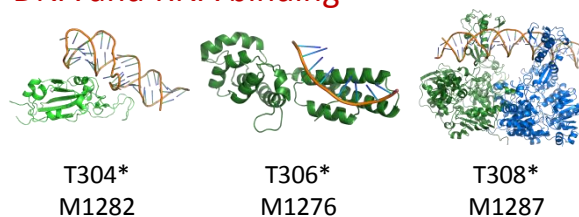


* No MassiveFold

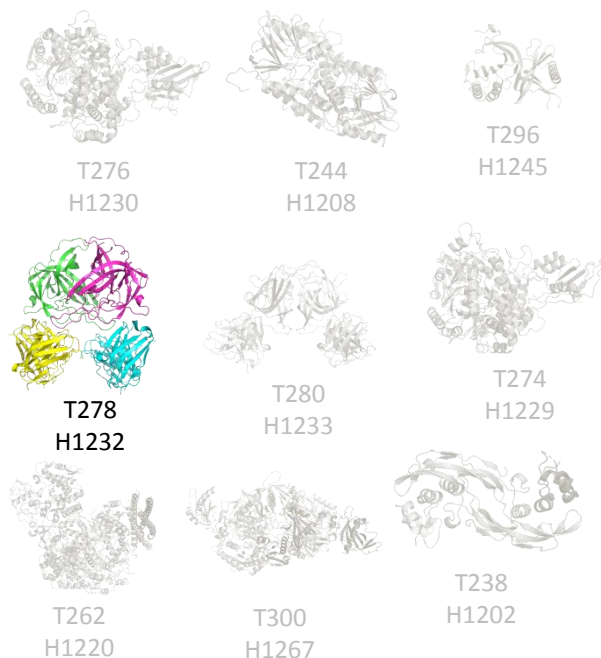
Conformers



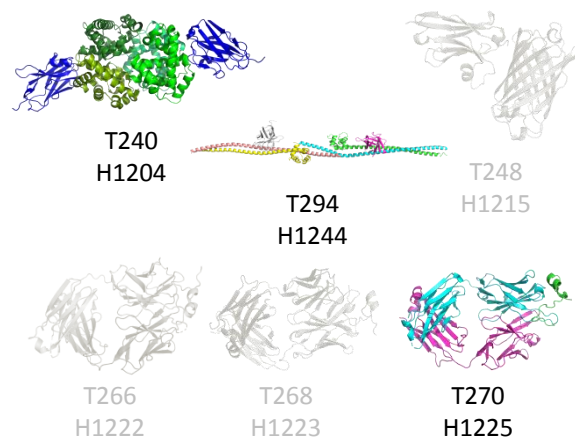
DNA and RNA binding



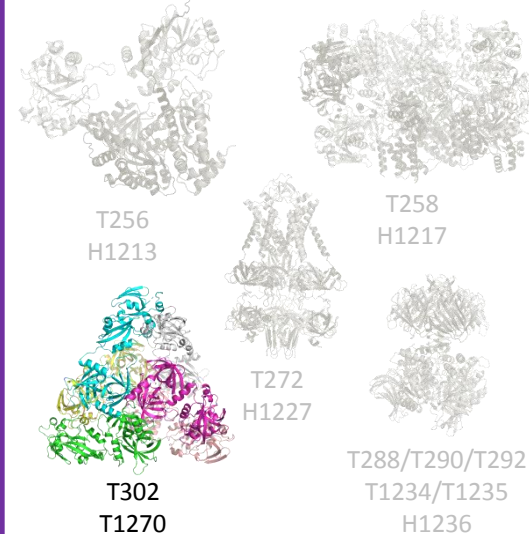
Hetero-oligomers



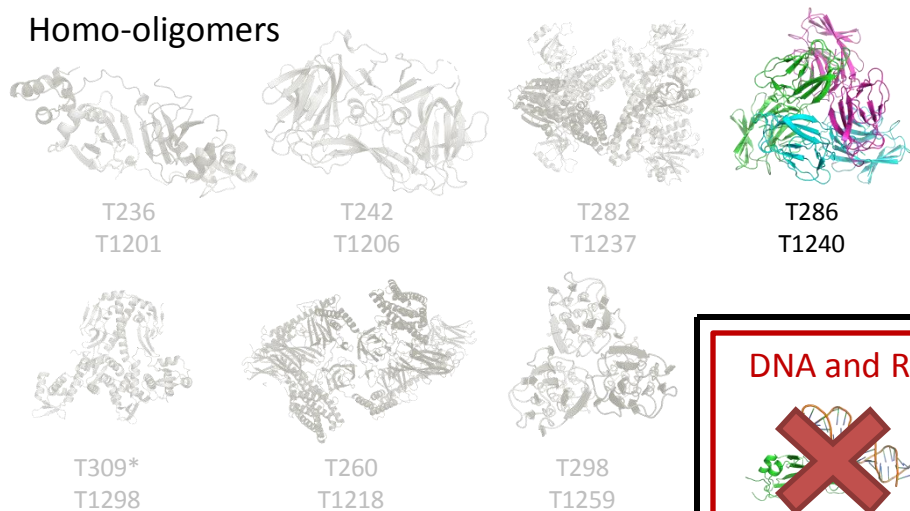
Antibodies and nanobodies



Large assemblies

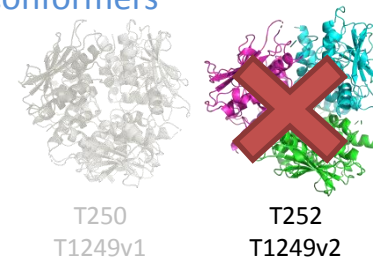


Homo-oligomers

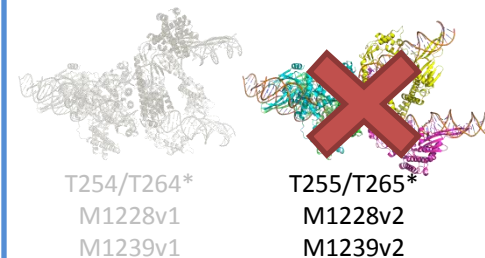
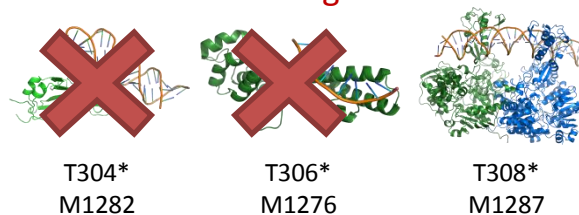


* No MassiveFold

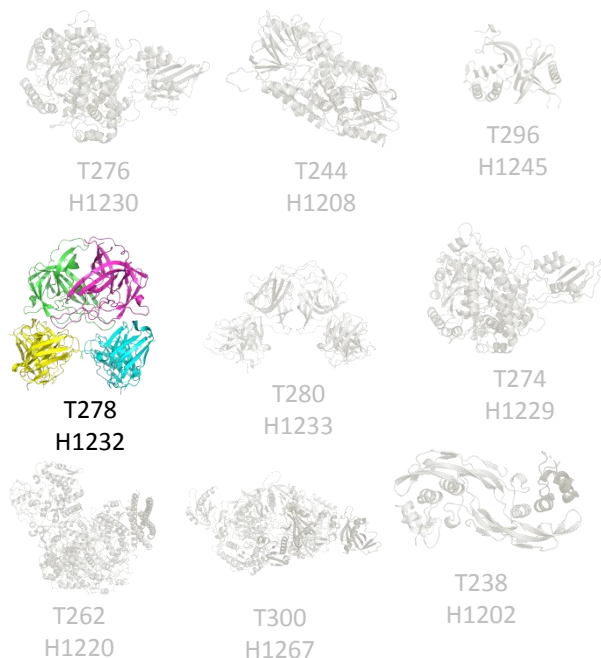
Conformers



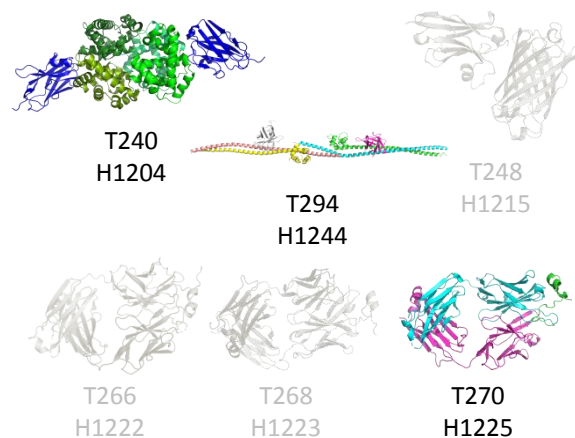
DNA and RNA binding



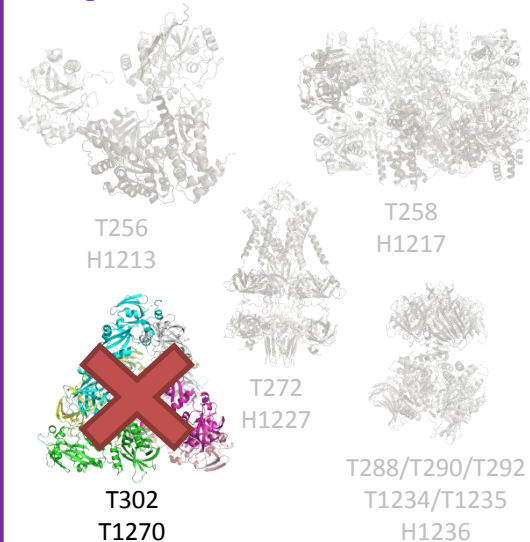
Hetero-oligomers



Antibodies and nanobodies



Large assemblies



(weak interface)

X-ray	-	A3
-------	---	----

	Stage1	Stage2
CASP	T1240	T2240
CAPRI	T286	T287



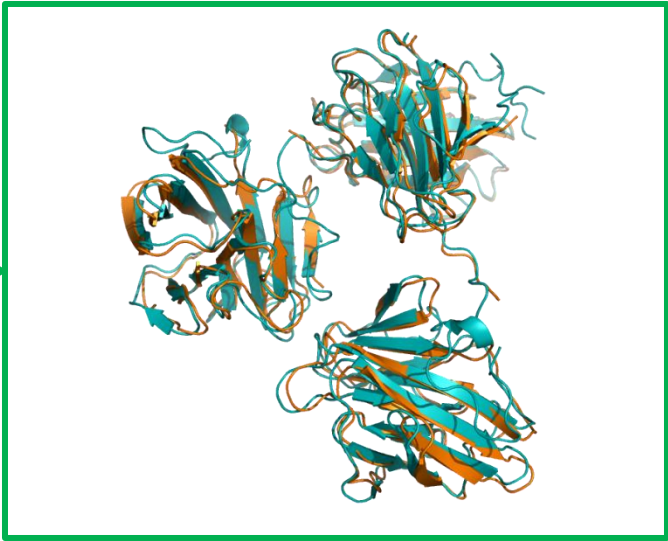
DIFFICULT

AlphaFold ranked_0	
rms	7.85 Å
ipTM	0.477

These are the *only* acceptable+ models

#Contacts	Interface	Chains	Area
135	1	A:B	900

INT1 – “homo”



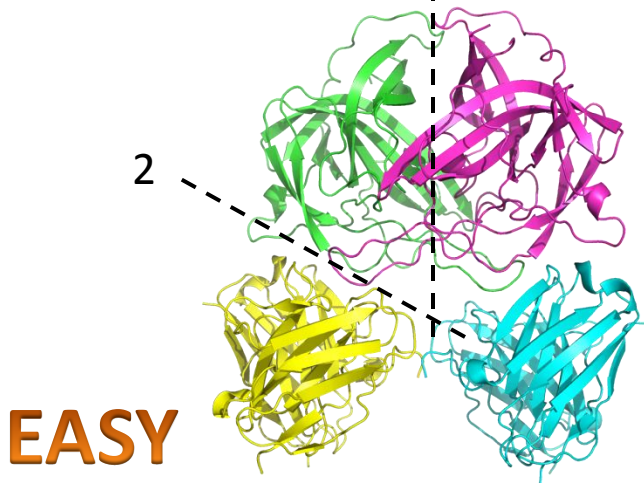
Group stage1	Model	Fnat	L-rms	i-rms	Classification
Wallner	1	0.88	1.53	1.04	Medium
S.Fernandez-Recio	9	0.79	7.72	2.54	Acceptable
MassiveFold	First acceptable model		Best quality model		Classification
16	#474		#512		Acceptable
Group stage2	Model	Fnat	L-rms	i-rms	Classification
Wallner	3	0.80	1.15	1.03	Medium
Fernandez-Recio	9	0.79	7.72	2.54	Acceptable

X-ray	-	A2B2
	Stage1	Stage2
CASP	H1232	H2232
CAPRI	T278	T279

Only A2 stable in solution, according to PISA

#Contacts	Interface	Chains	Area
200	1	A:B	1285
334	2	AB:C	450

INT1 – “homo”, “dimer”
INT2 – “hetero”



EASY

Top-1

T278/H1232	CAPRI (20)	CASP (72)	Scorers (13)	
Interface 1	20/18***/2**	67/60***/7**	10/9***/1**	
Interface 2	0	1	0	
T279/H2232	CAPRI (18)	CASP (58)	MassiveFold	
Interface 1	18/17***	57/52***	4619***	high
Interface 2	0	1	66	incorrect

AlphaFold ranked_0	
rms	x Å
ipTM	0.781

These are the *only* acceptable+ models

Group stage1	Model	Fnat	L-rms	i-rms	Classification
Smg-ulaval	2	0.51	6.35	1.13	Medium
Gray, Kihara, Zou, MDCKPP, Pierce, FALCON2					Acceptable
MassiveFold	First acceptable model		Best quality model		Classification
66	#2509		#3531		Acceptable
Group stage2	Model	Fnat	L-rms	i-rms	Classification
Plmfold	4	0.39	14.84	1.94	Medium
FALCON2, plmfold, Zou, MILLISECONDS					Acceptable

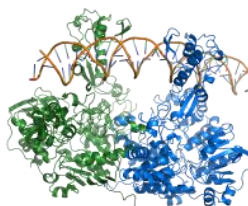
DNA and RNA binding



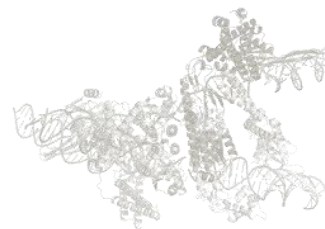
T304*
M1282



T306*
M1276



T308*
M1287



T254/T264*
M1228v1
M1239v1



T255/T265*
M1228v2
M1239v2

Predictors

Bhattacharya

Fernandez-Recio

ALPHAFOLD3-SERVER

B-LAB

GeneSilico

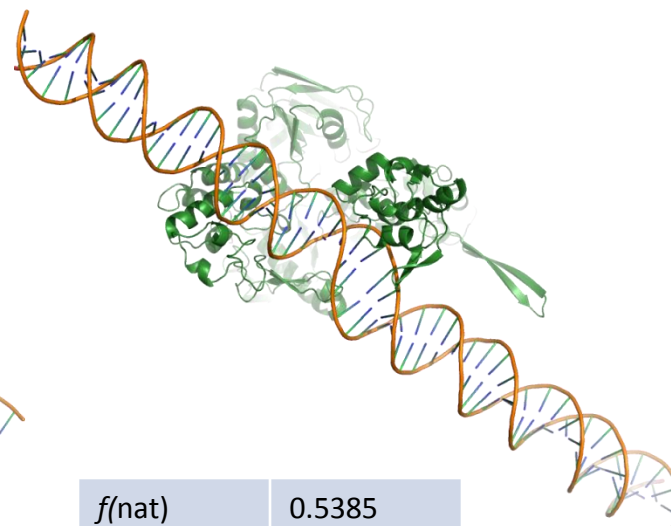
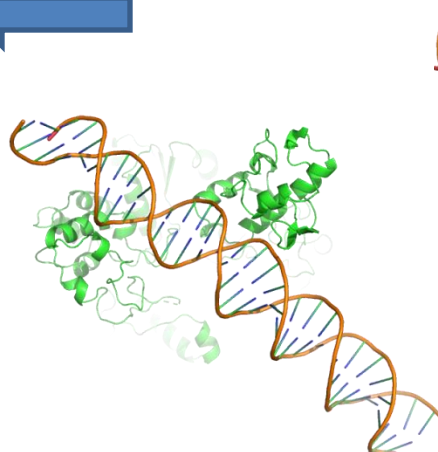
Diff

Scorers

Fernandez-Recio

Oliva

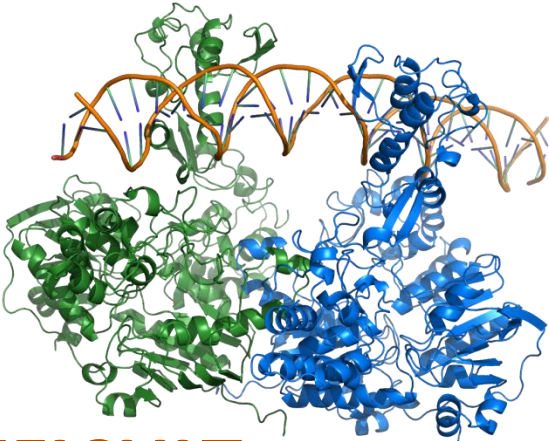
Kihara / LZERD



$f(\text{nat})$	0.5385
$f(\text{nonnat})$	0.8454
$i\text{-rms}$	3.8529

EM		
----	--	--

	Stage1	Stage2
CASP	M1287	-
CAPRI	T308	-



DIFFICULT

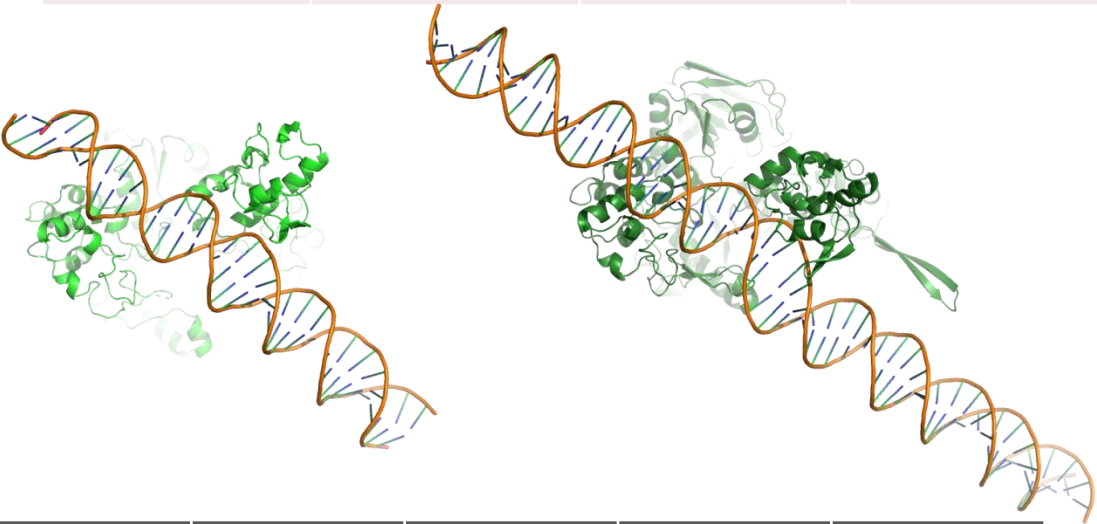
AlphaFold ranked_0	
rms	x Å
ipTM	NA

#Contacts	Interface	Chains	Area
227	1	A:C	1565
164	2	A:DNA	420

INT1 – “homo”, “dimer”
INT2 – “DNA/RNA”

Top-1

T308/M1287	CAPRI (16)	CASP (29,28)	Scorers (12,11)
Interface 1	14/2**	19/3**	12
Interface 2	0	1	0



Group stage1	Model	Fnat	L-rms	i-rms	Classification
Bhattacharya	5	0.54	16.55	3.85	Acceptable
Diff, Fernandez-Recio, B-LAB , GeneSilico, AlphaFold3-server					Acceptable
<i>S.Fernandez-Recio, S.Kihara, S.LZERD, S.Oliva</i>					Acceptable

102 acceptable in
the MassiveFold set
First: #120
Best: #173

High

Kozakov/Vajda

Acceptable

Cheng, Pierce, Zou

Fernandez-Recio

HYU-MLLab

LupengKong

MRAFold

MULTICOM (all variants)

OmniFold

YANG-MULTIMER, Yang, Yang-Server

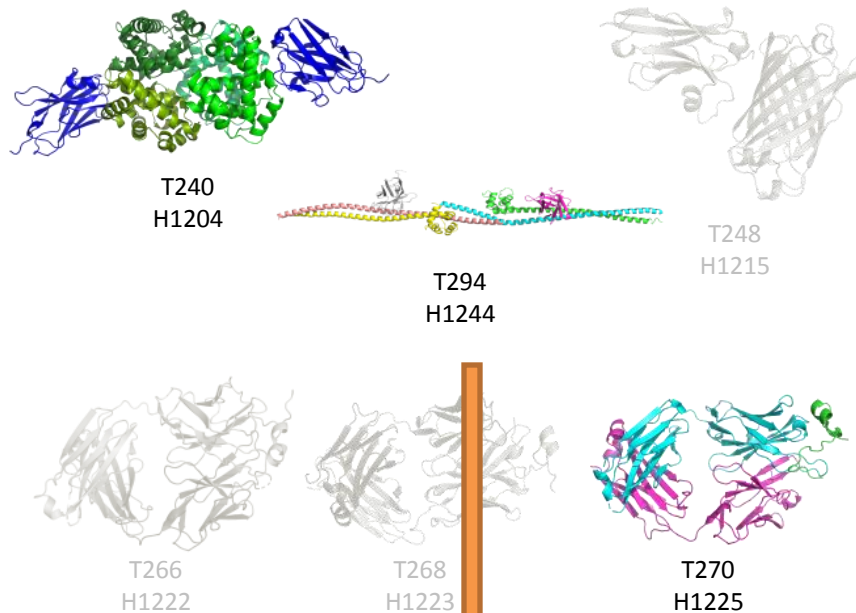
MQA-SERVER

CSSB-Faker

FALCON2

Zou

Antibodies and nanobodies



No acceptable
solutions for
T270/H1225

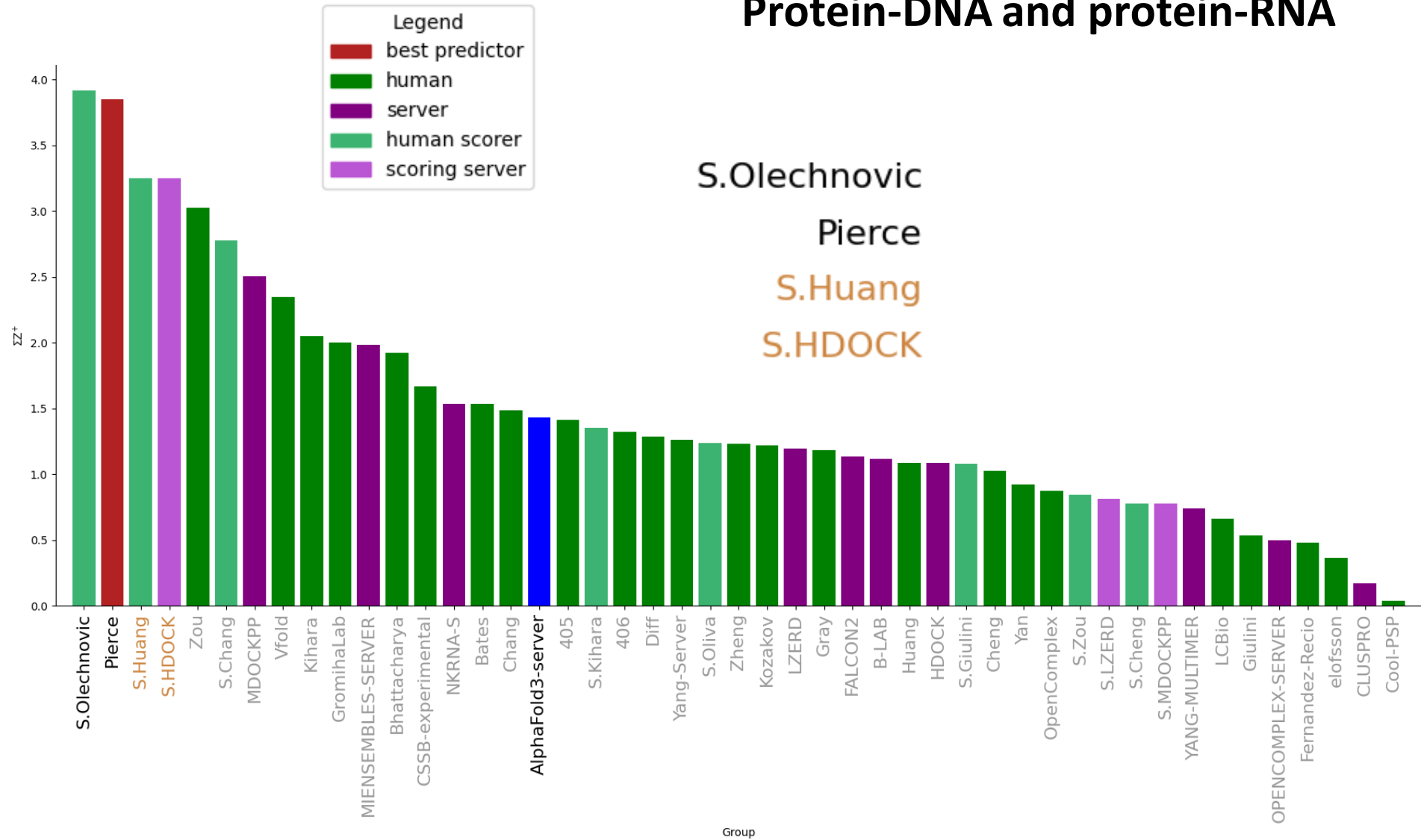
25 acceptable in the
MassiveFold set
First: #249
Best: #4902

No acceptable
solutions for
T294/H1244

No good models in
the MassiveFold set

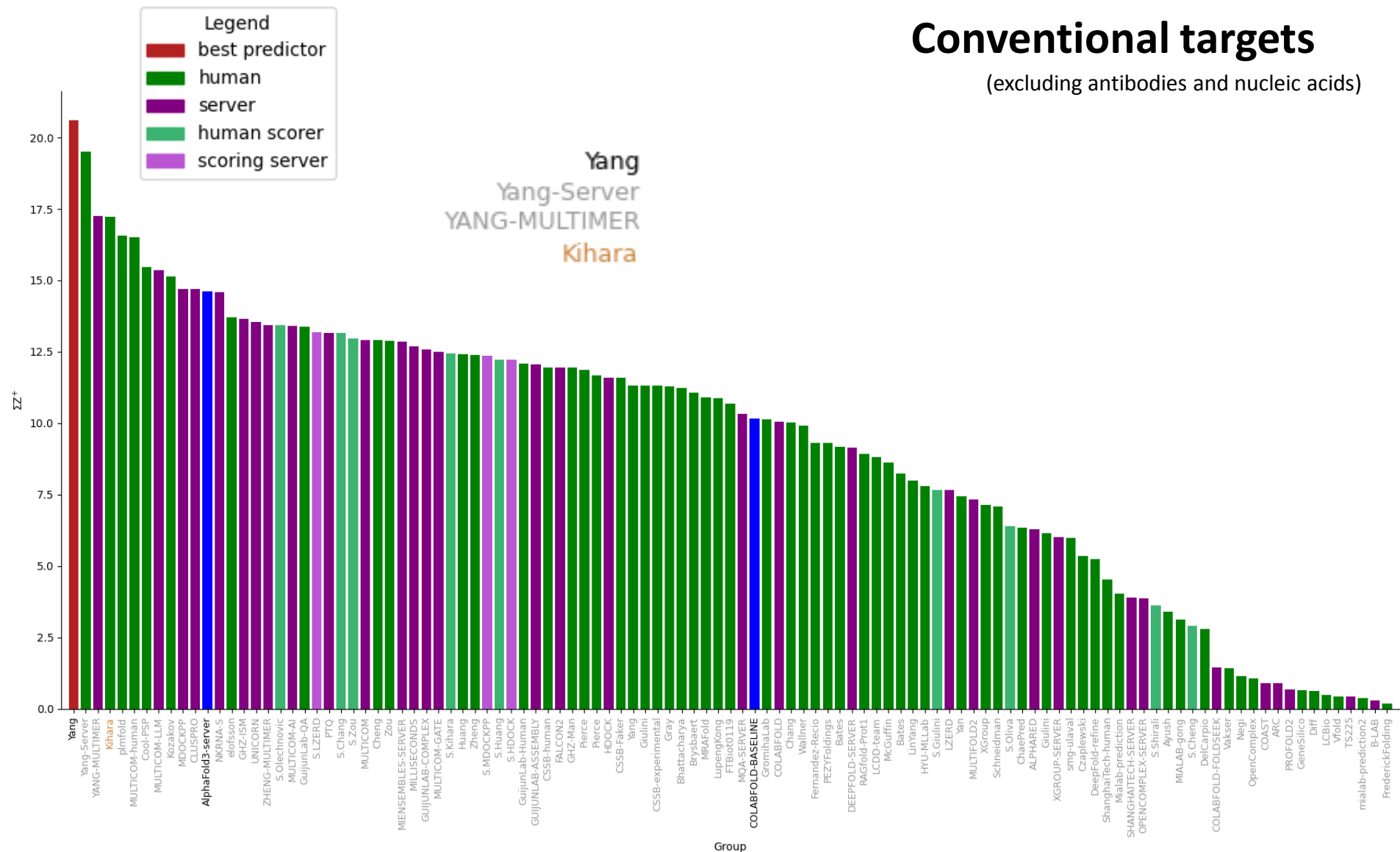
PERFORMANCE IN CATEGORIES

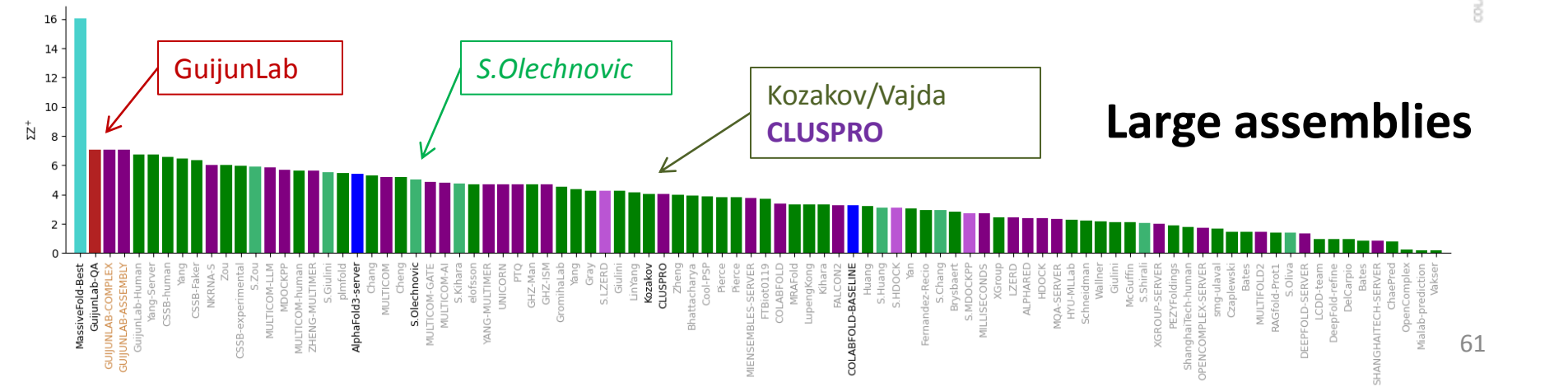
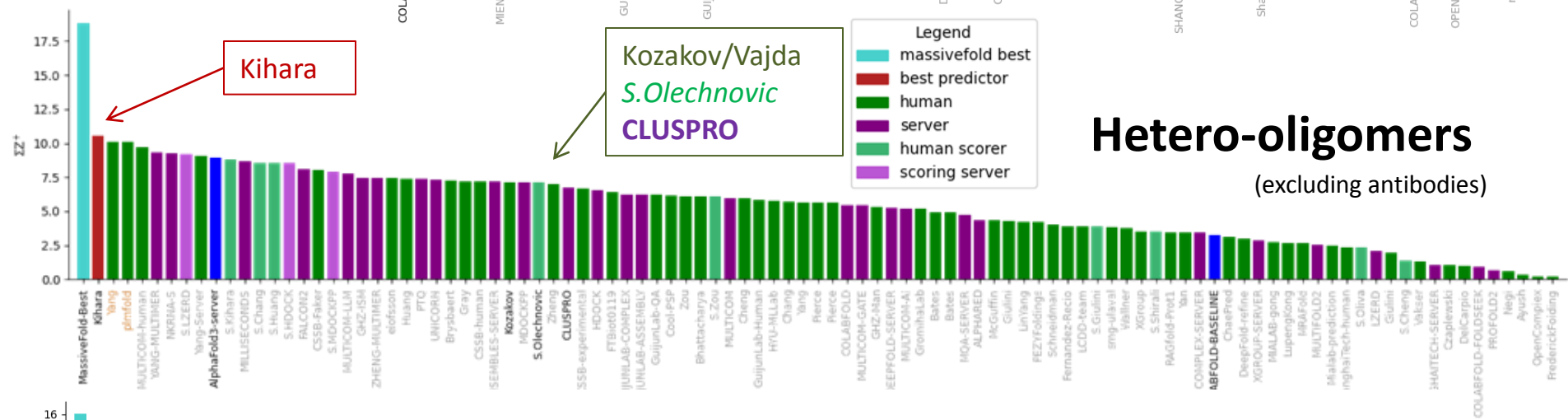
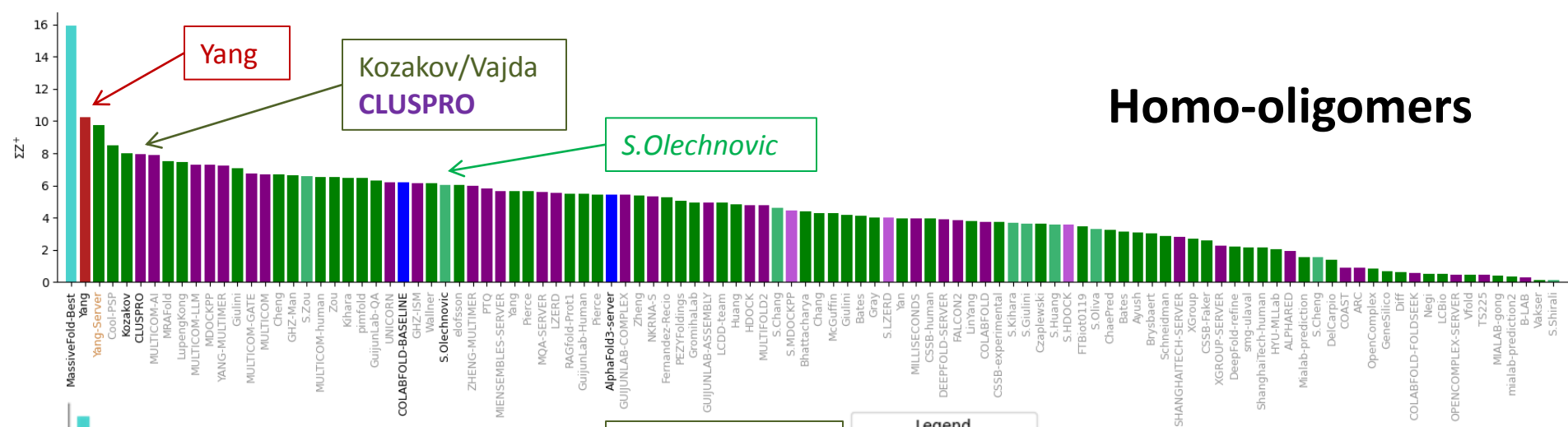
Protein-DNA and protein-RNA

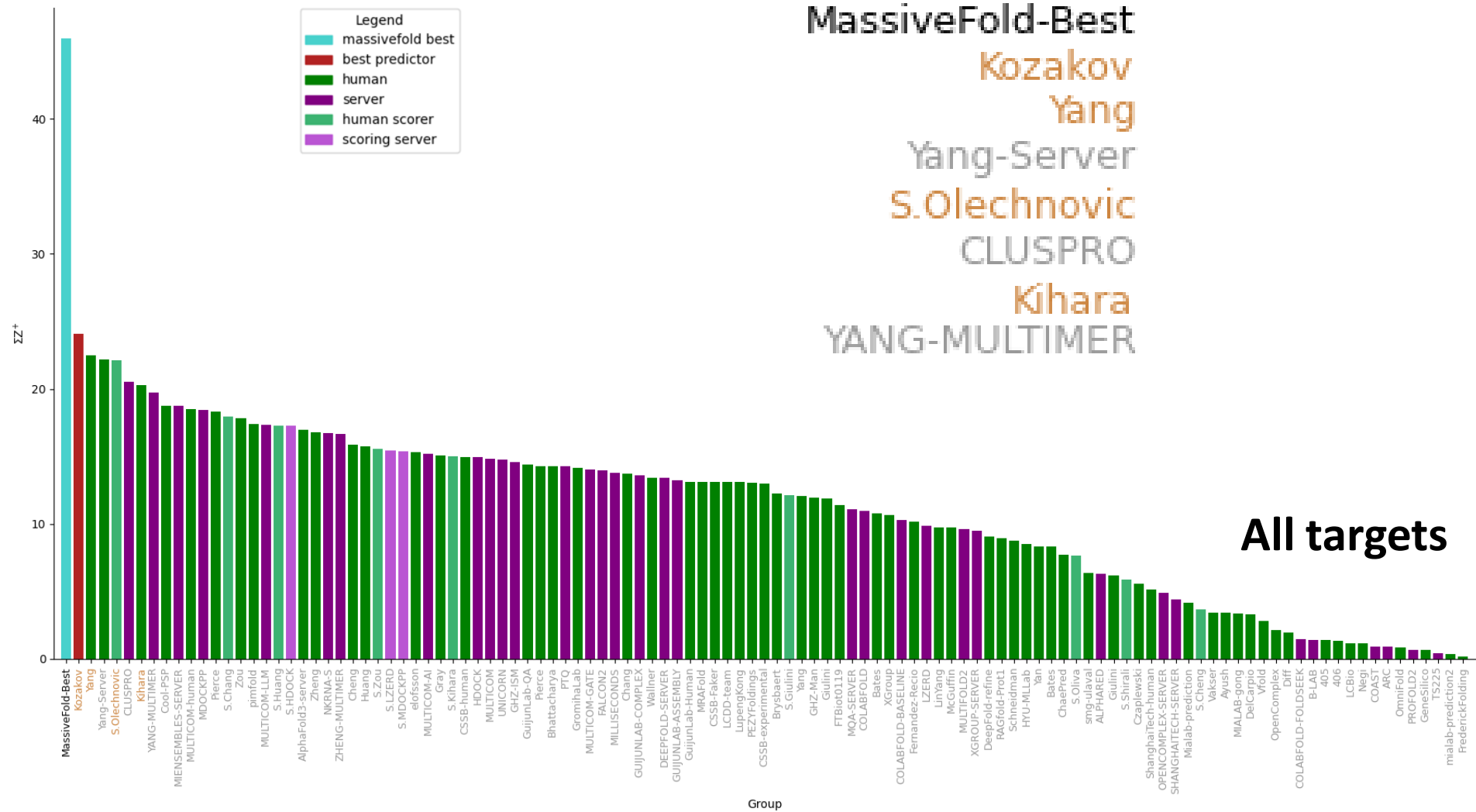


Conventional targets

(excluding antibodies and nucleic acids)





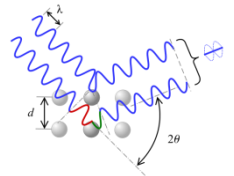
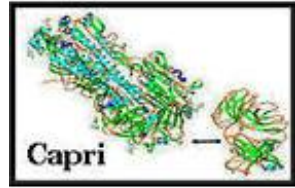


Conclusions

- PRO'S:
 - This year's session was a big success
 - Flat distribution of scores; many predictors producing good models
 - The best predictors find good models for 75% of targets
 - Kihara, Cheng, Kozakov/Vajda, Huang⁺, Zou
 - Fewer targets found but of better quality
 - MULTICOM, Zheng, Brysbaert, GuijunLab
 - Significant progress in the prediction of antibody and nanobody binding
 - Best performance by Kozakov/Vajda
 - In terms of relative prediction performance, some groups stand out:
 - Kozakov/Vajda⁺, Yang⁺, Kihara
 - scorer Olechnovic
 - LZERD for stage2 vs stage1, and for scoring
- CON'S:
 - Alternate conformers were **not** found
 - Nucleic acids remain problematic; DNA is easier than RNA
 - Scoring needs more development
 - MassiveFold “best” outperforms everybody, especially on “conventional” targets
 - Scorers recognize the *good* models, but not the *best* ones

Acknowledgements

- CAPRI community
- CASP community
- CASP organizers
- The MassiveFold team & IFB, IDRIS
- The experimentalists
- DeepMind

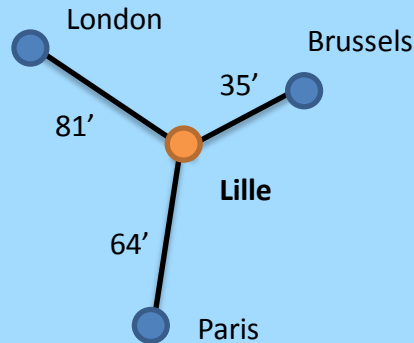




Computational Biology: Biomolecular Interactions and Dynamics

Institute for Structural and Functional Glycobiology

- **Protein interaction** and regulatory networks
- **Statistical physics of biomolecular interactions**
- **Structural biology of protein-carbohydrate interactions**
- **Computational modeling and dynamics of biomolecular systems**
- **The impact of evolution on protein-protein interaction and vice versa**



CECAM

